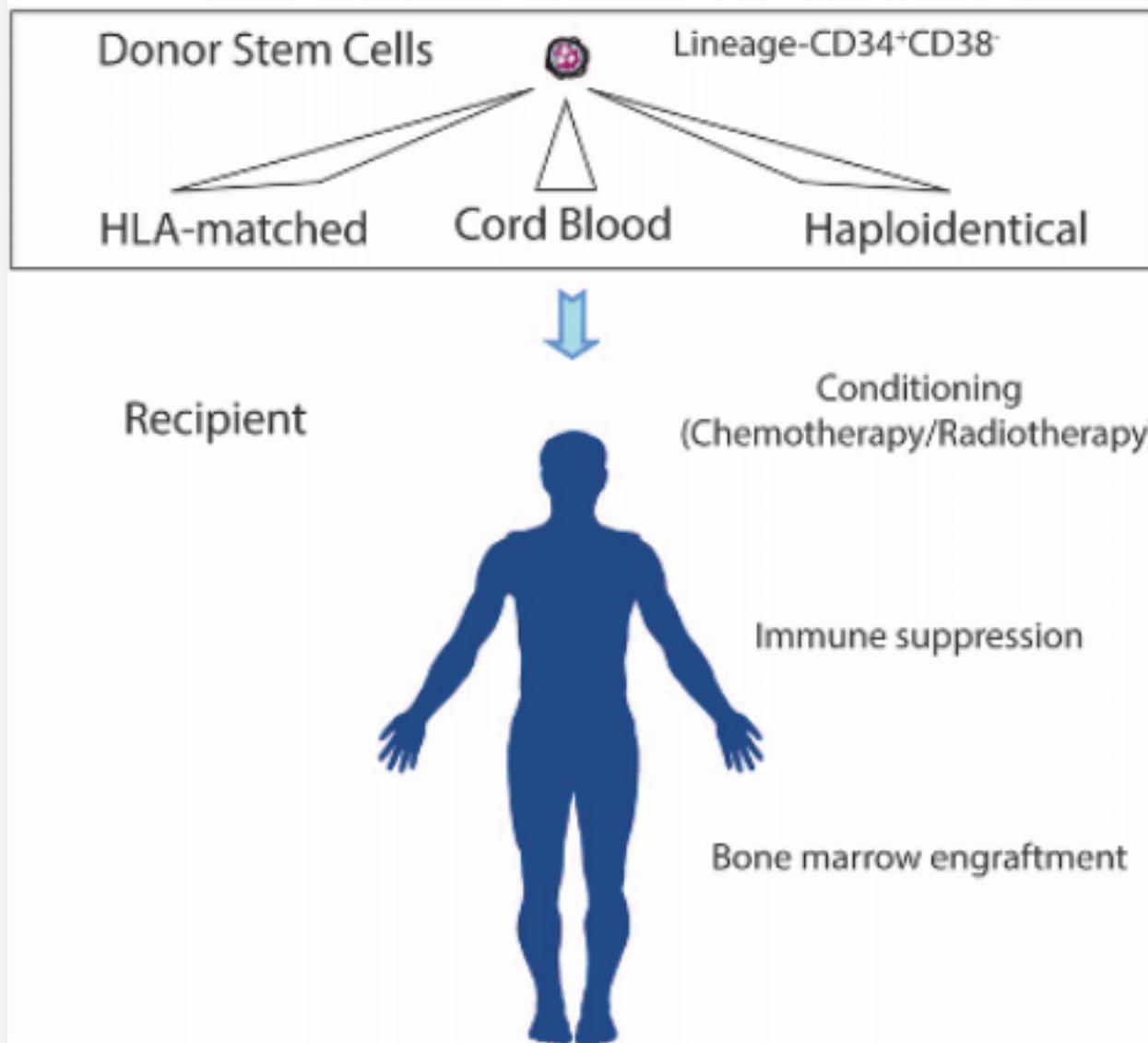


شناسایی اهدا کننده مناسب سلولهای بنیادی در پیوند نقایص ایمنی
اولیه و
گزارشی از مرکز پذیره نویسی اهدای سلولهای بنیادی مرکز تحقیقات
ایمونولوژی، آسم و آلرژی

Maryam Nourizadeh, PhD in Medical Immunology
Assistant Prof.,
Immunology, Asthma and Allergy Research Institute (IAARI)
Tehran University of Medical Sciences (TUMS)

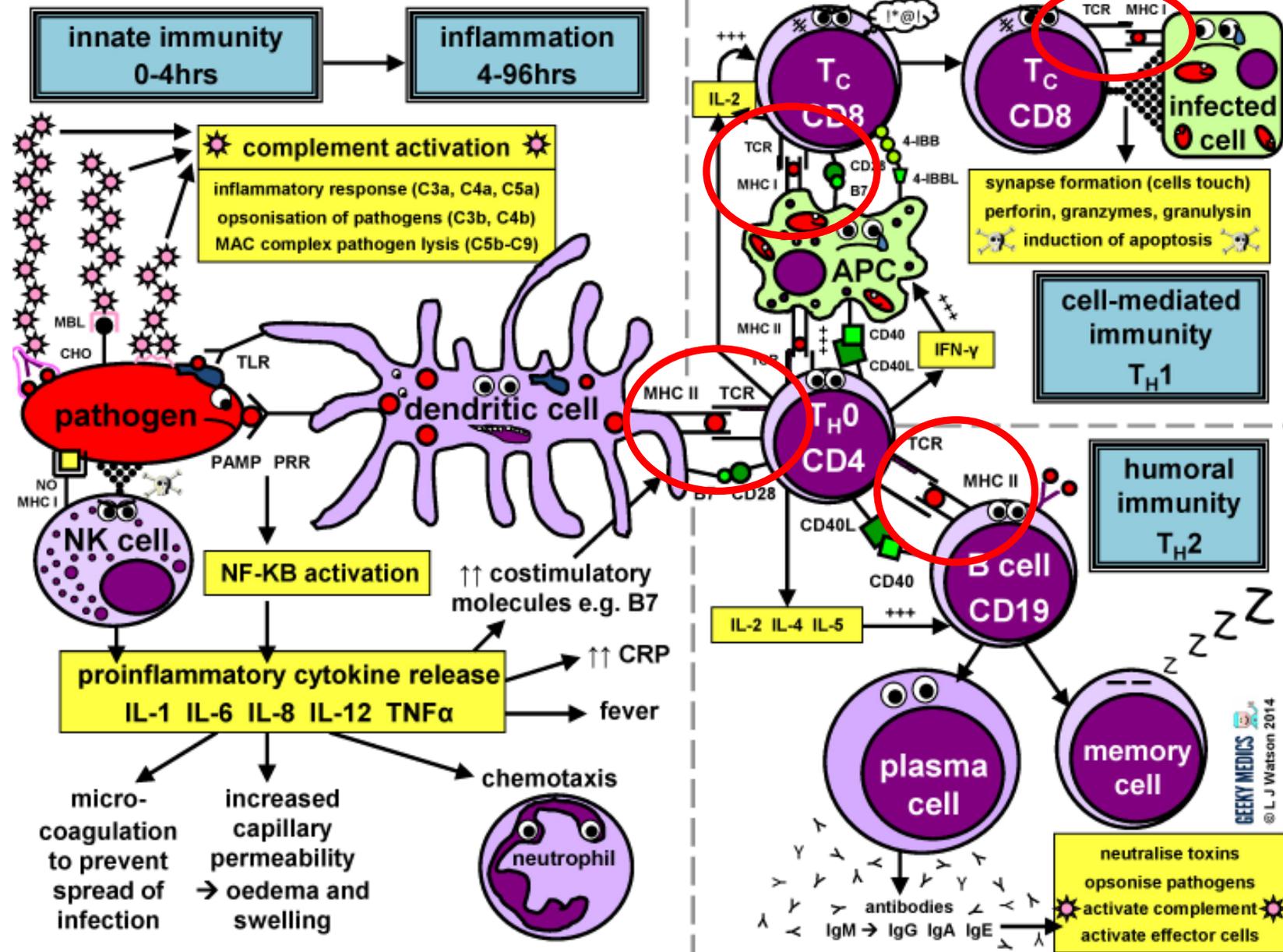
7 MARCH 2024

Allogeneic Stem Cell Transplant Therapy

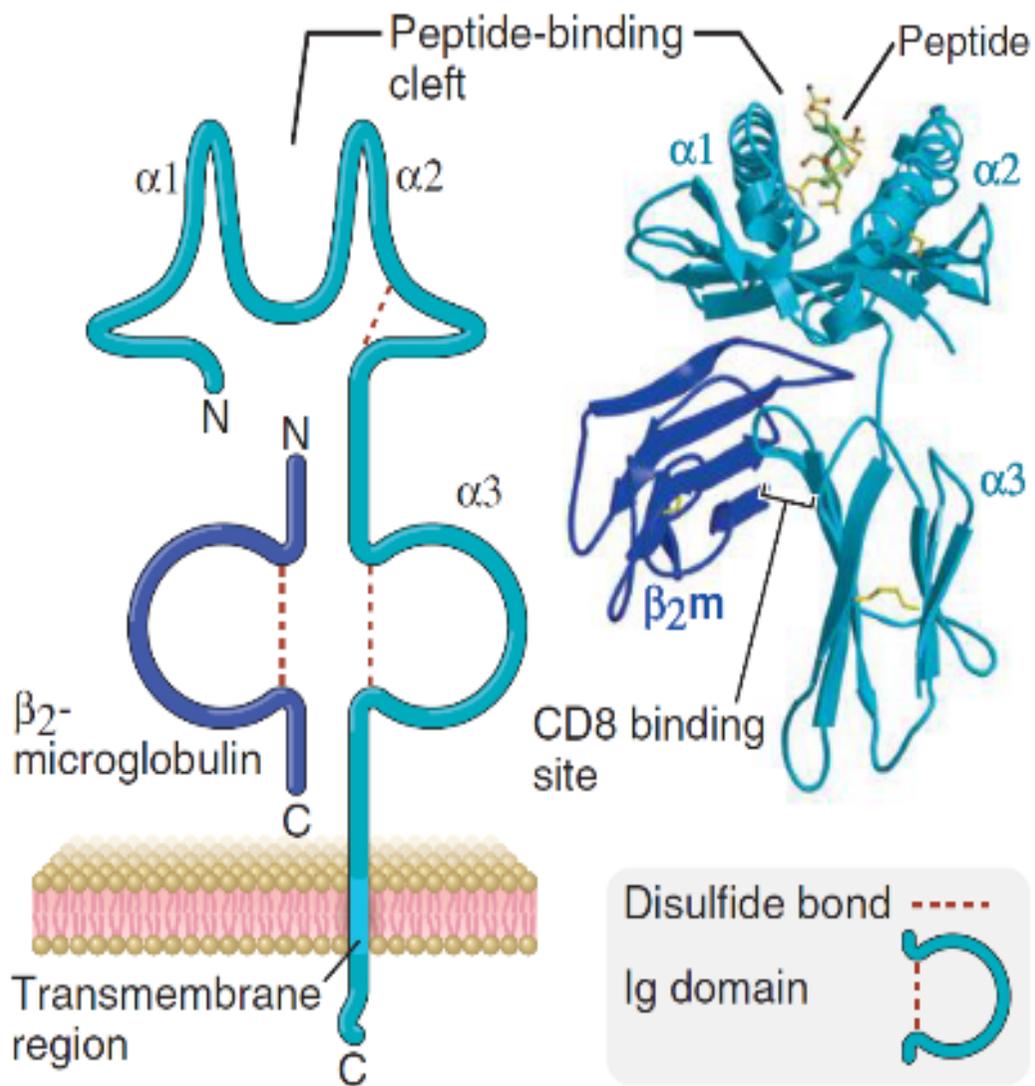


What is HLA? What is it for?

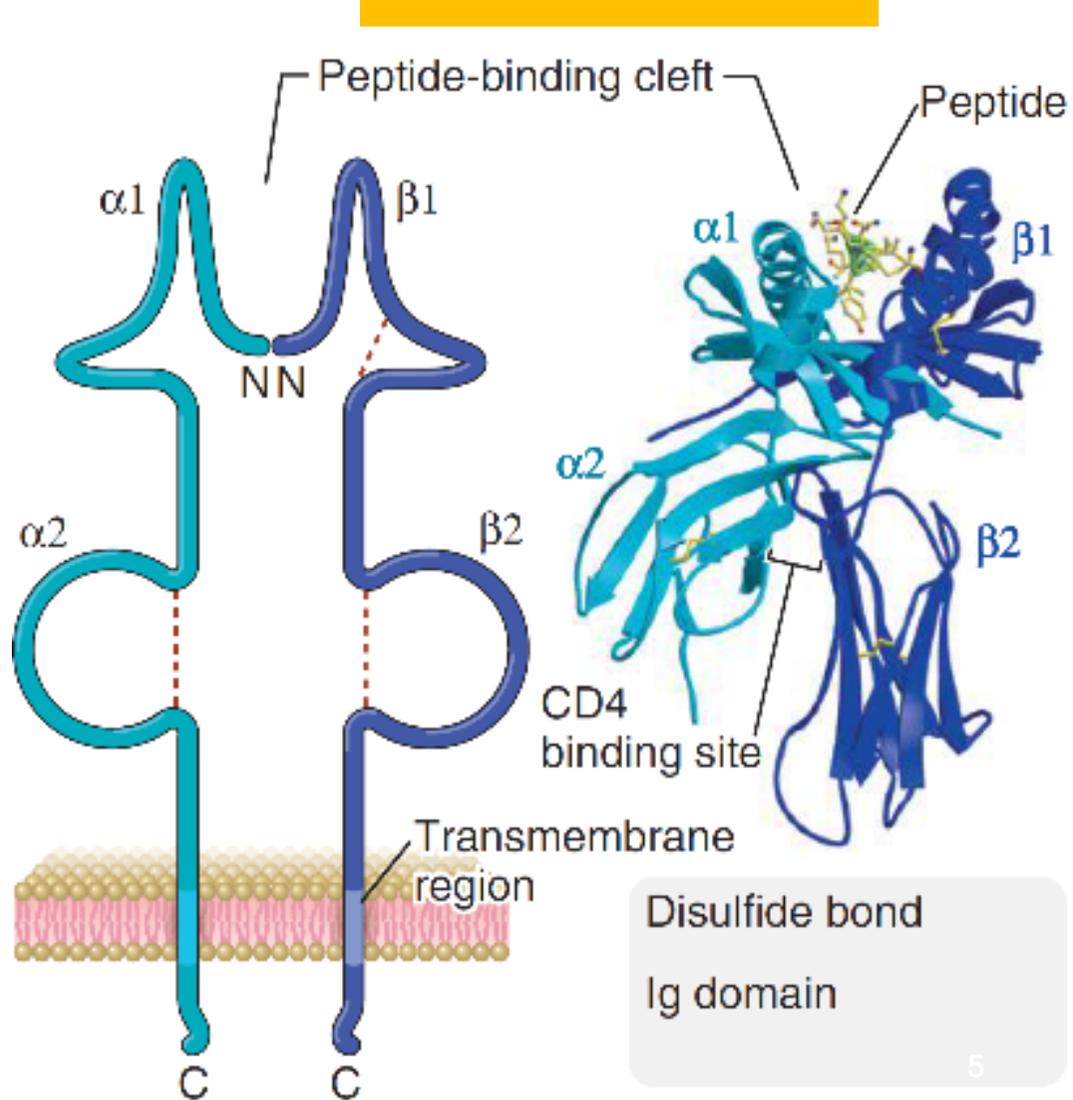
- Human Leukocyte Antigen
- Discovered: in mice (1937), humans (1954)
- Function: to present peptides to T cells, thus allowing elimination of foreign particles and recognition of self (so in transplants this has to be modulated)



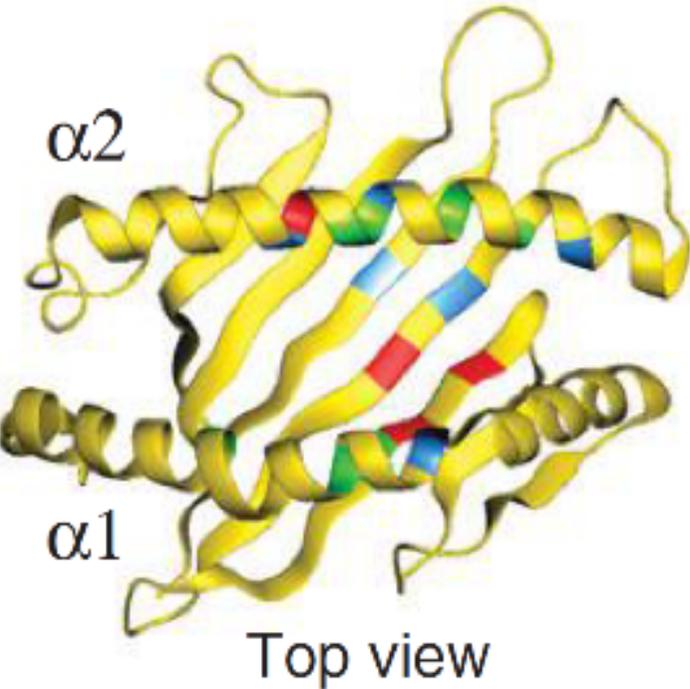
MHC class I expressing cells



MHC class II expressing cells

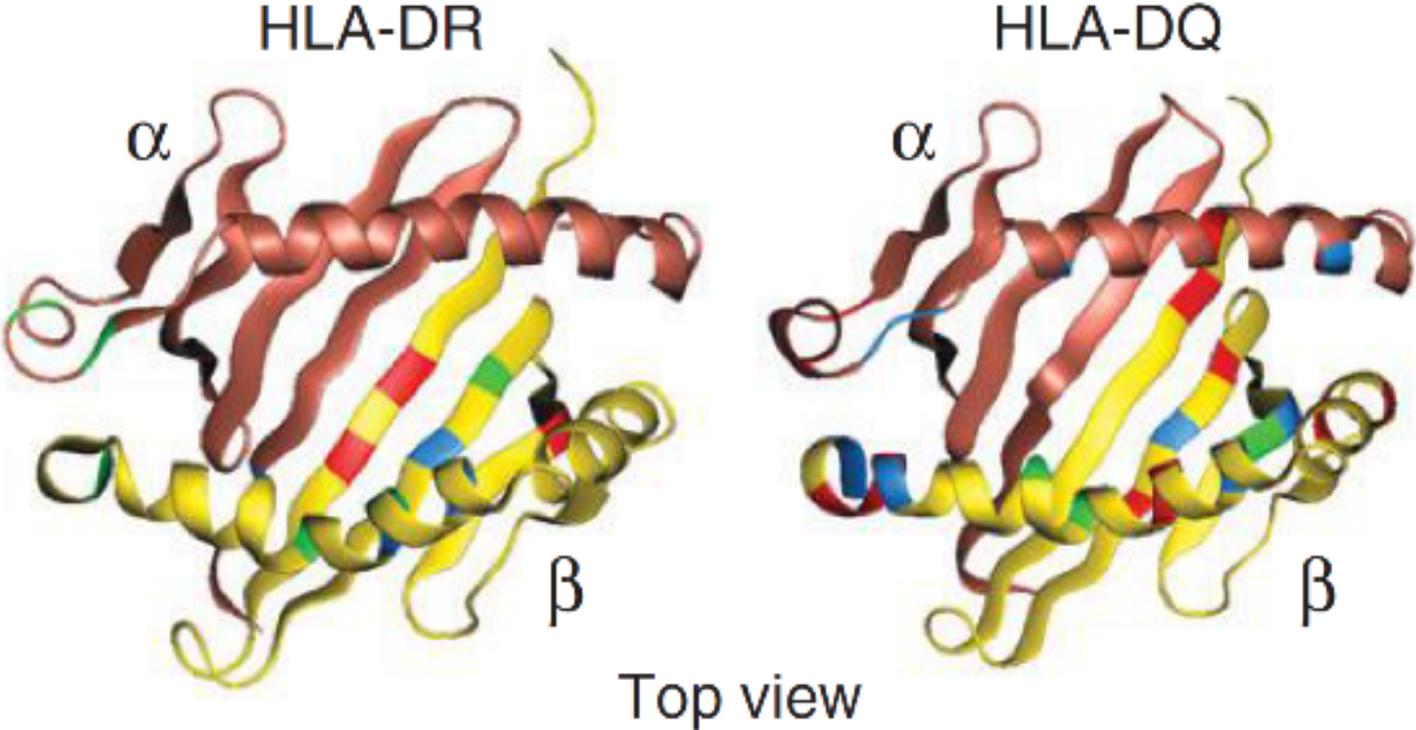


HLA class I



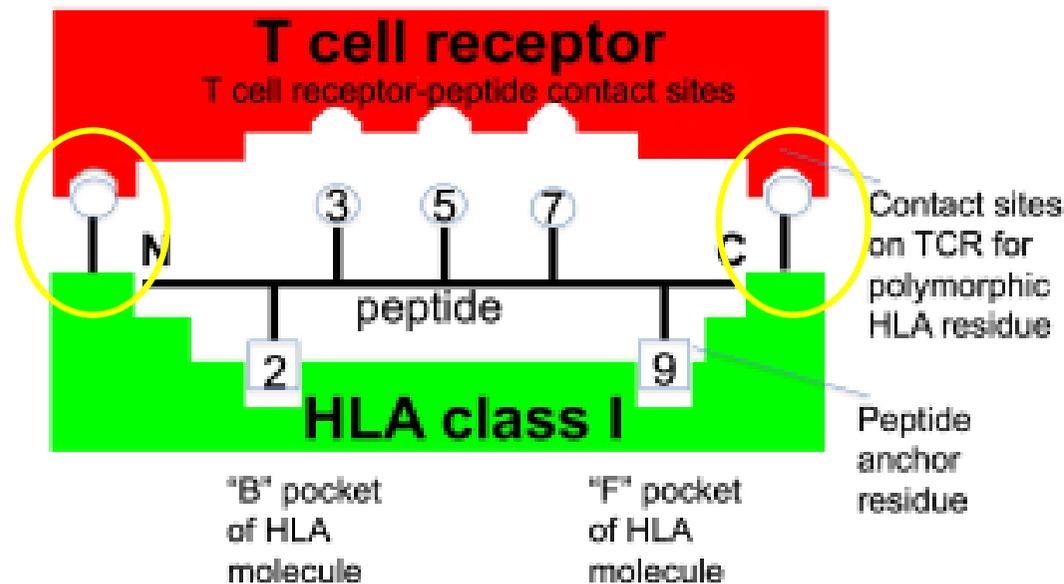
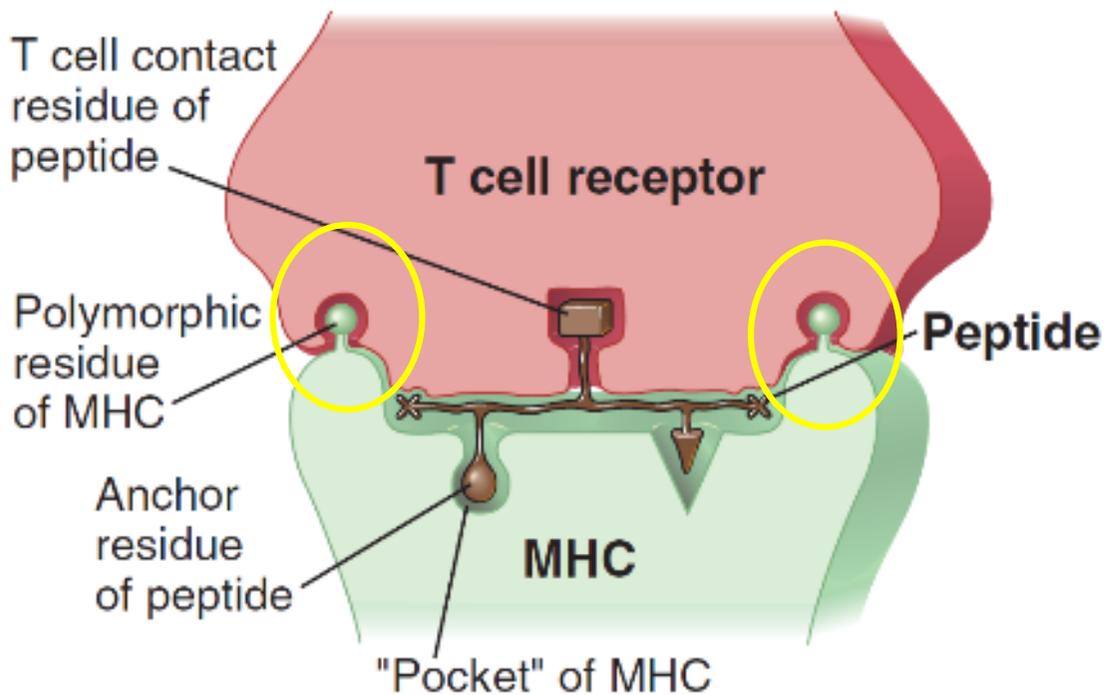
154'818 peptides
length 8 to 10 aa

HLA class II



> 2000 peptides
length 12-24 aa

- MHC is the most highly polymorphic gene system in the body and hence in the population.
- This extensive polymorphism of MHC genes makes it very unlikely that two random individuals will express identical sets of MHC molecules.
- This polymorphism is the basis of rapid graft rejection between genetically different individuals.



Chromosome 6

Long arm

Short arm

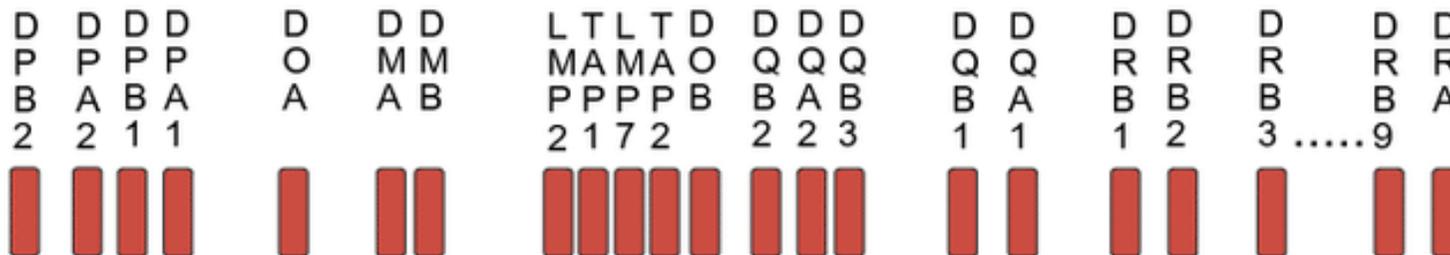
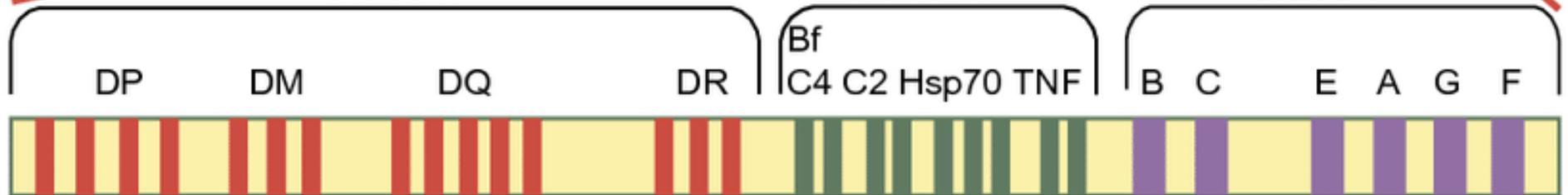


HLA region
6p21.1-21.3

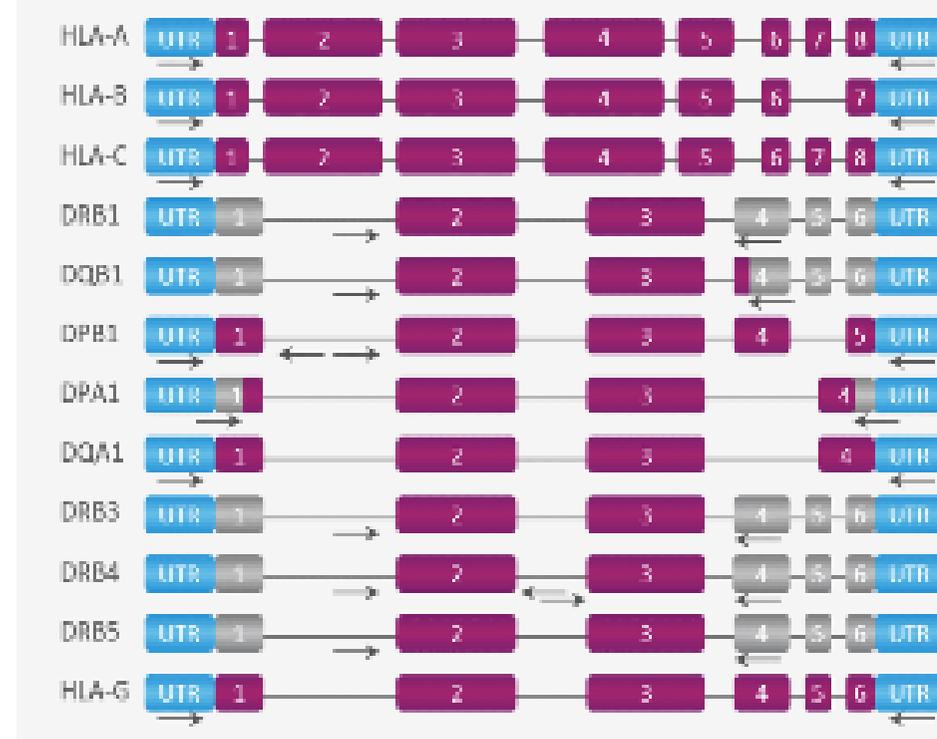
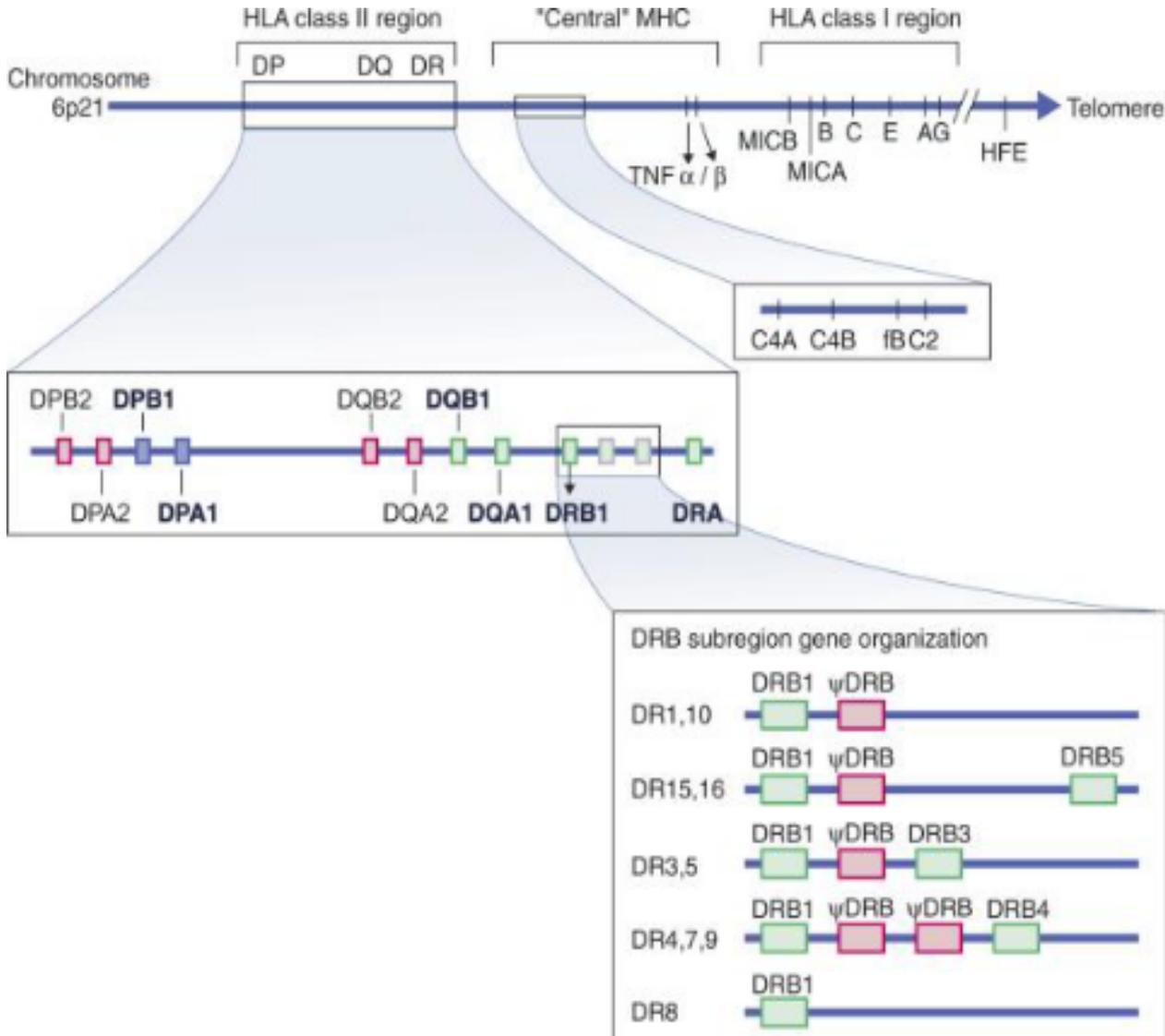
Class II

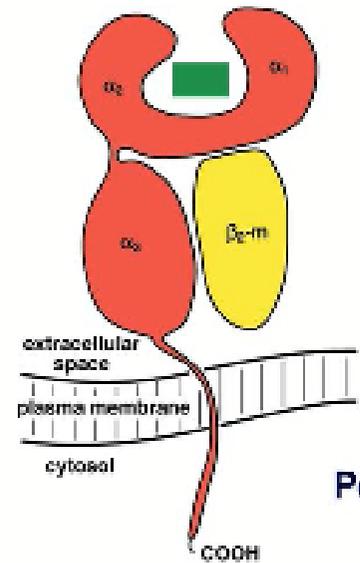
Class III

Class I



Human MHC Gene Loci

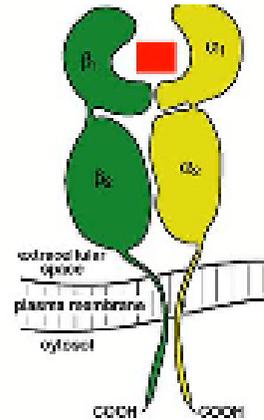
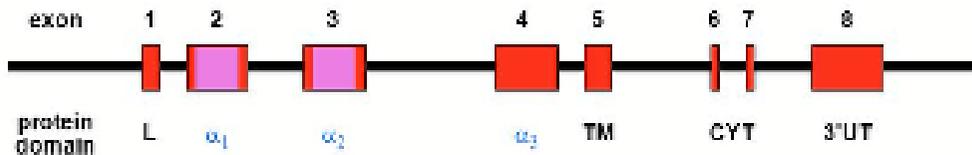




HLA Class I

- found on all nucleated cells

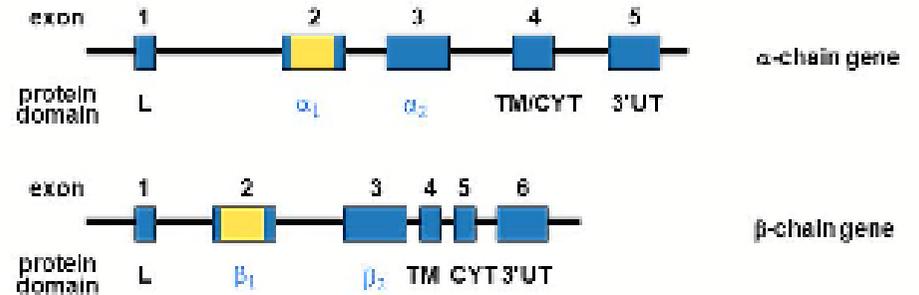
Polymorphism located in exons 2 & 3



HLA Class II

- restricted to cells of the immune system

Polymorphism located in exon 2



Welcome to IPD

Jan 2023, build 72

IPD was developed in 2003 to provide a centralised system for the study of polymorphism in genes of the immune system. The IPD project was established by the HLA Informatics Group of the Anthony Nolan Research Institute in close collaboration with the European Bioinformatics Institute.



IPD Resources

[Submission](#)

[RFST APT](#)

[Citations](#)

[Contact](#)

[Support](#)

Version report - 3.55 (2024-01)

4 Versions in a year
(Jan, April, July, Oct)

The following sequences have been submitted to the Nomenclature Committee since the previous release and have been assigned official allele designations.

For a complete release list, please visit the [release list page](#).

New alleles

Allele	Accession	Assigned Date	EMBL/GenBank	Comments
A*01:01:01:116	HLA39086	2023-12	DH247664	Genomic Sequence
A*01:01:01:117	HLA39048	2023-12	DH664359	Genomic Sequence
A*01:01:01:153	HLA39122	2023-12	DH839877	Genomic Sequence

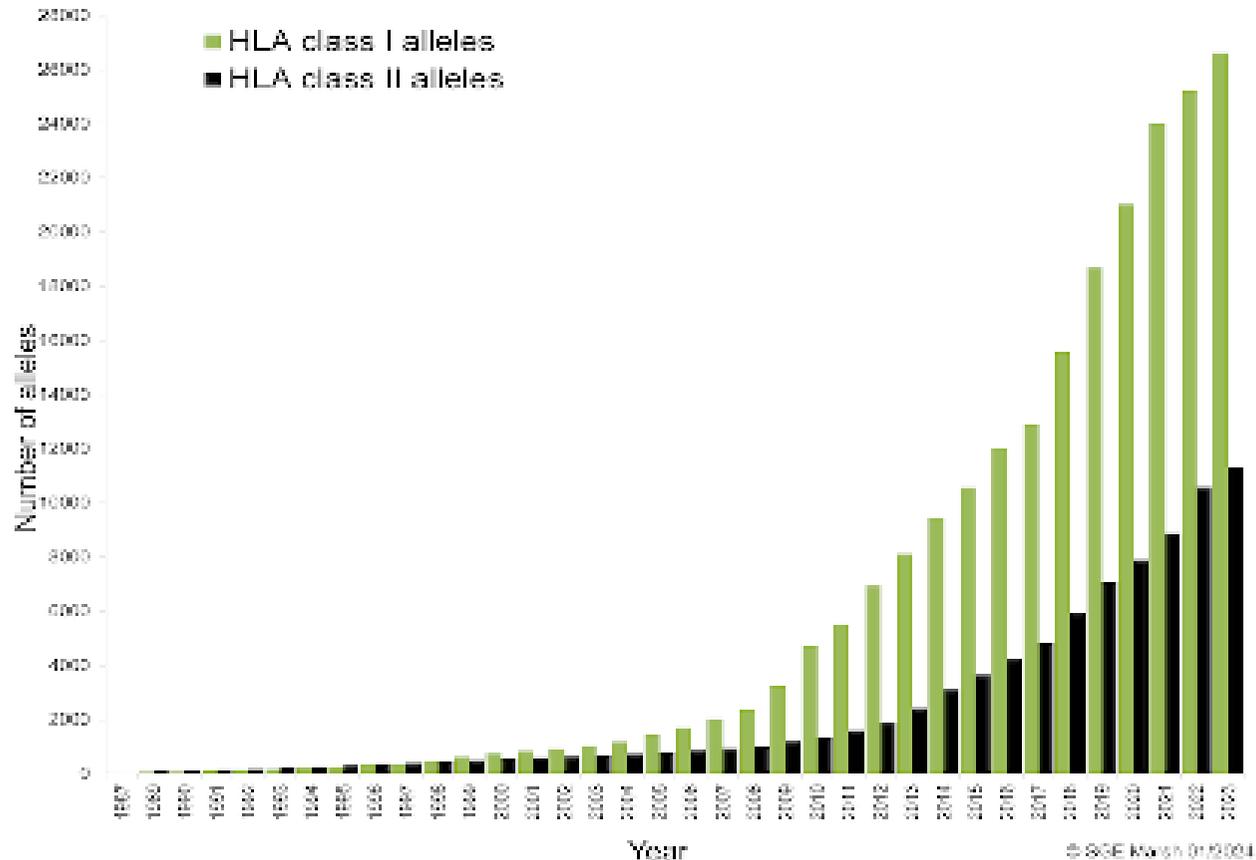
Summary

New alleles (499)
Modified alleles (128)
Suffix changes (3)
Deleted alleles (1)

Database Growth

The IPD-IMGT/HLA Database was first released in 1998 with 964 alleles. The latest version has 38909 distinct allelic variants.

The graph below indicates the numbers of alleles named and held in the IPD-IMGT/HLA Database since 1997 up to 2024-01-16.



HLA Alleles Assigned as of January 2024

HLA class I						
Gene	<i>A</i>	<i>B</i>	<i>C</i>	<i>E</i>	<i>F</i>	<i>G</i>
Alleles	8098	9656	8084	352	91	158
Proteins	4741	5745	4470	140	17	48
Nulls	420	339	362	10	2	6

HLA Alleles Assigned as of January 2024

HLA class II														
Gene	<i>DRA</i>	<i>DRB</i>	<i>DQA1</i>	<i>DQA2</i>	<i>DQB1</i>	<i>DQB2</i>	<i>DPA1</i>	<i>DPA2</i>	<i>DPB1</i>	<i>DPB2</i>	<i>DMA</i>	<i>DMB</i>	<i>DOA</i>	<i>DOB</i>
Alleles	67	4581	722	42	2510	41	639	6	2486	7	59	82	93	63
Proteins	15	3019	351	11	1524	9	311	0	1447	0	9	9	14	16
Nulls	0	200	21	0	111	1	26	0	130	0	0	0	1	0

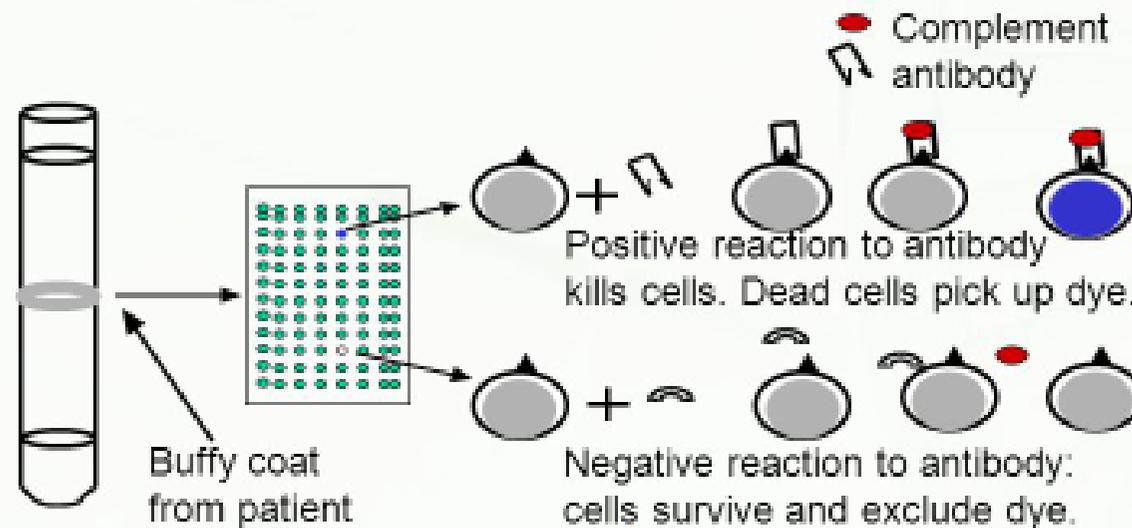
HLA class II - DRB Alleles										
Gene	<i>DRB1</i>	<i>DRB2</i>	<i>DRB3</i>	<i>DRB4</i>	<i>DRB5</i>	<i>DRB6</i>	<i>DRB7</i>	<i>DRB8</i>	<i>DRB9</i>	
Alleles	3628	1	488	250	201	4	2	1	6	
Proteins	2343	0	369	157	150	0	0	0	0	
Nulls	124	0	24	27	25	0	0	0	0	

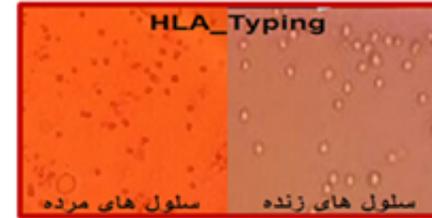
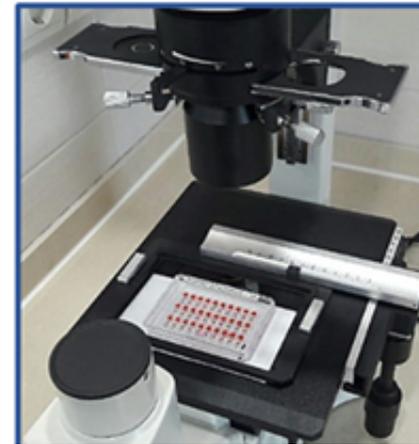
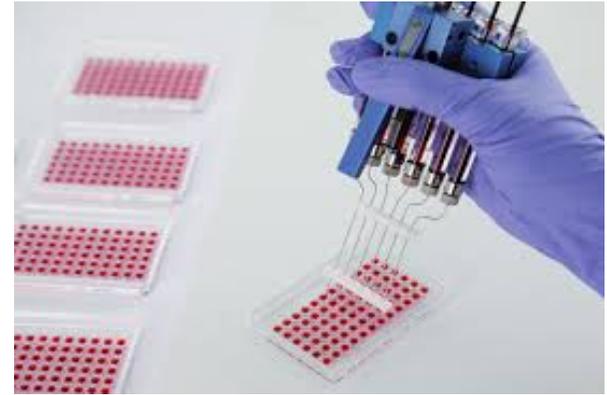
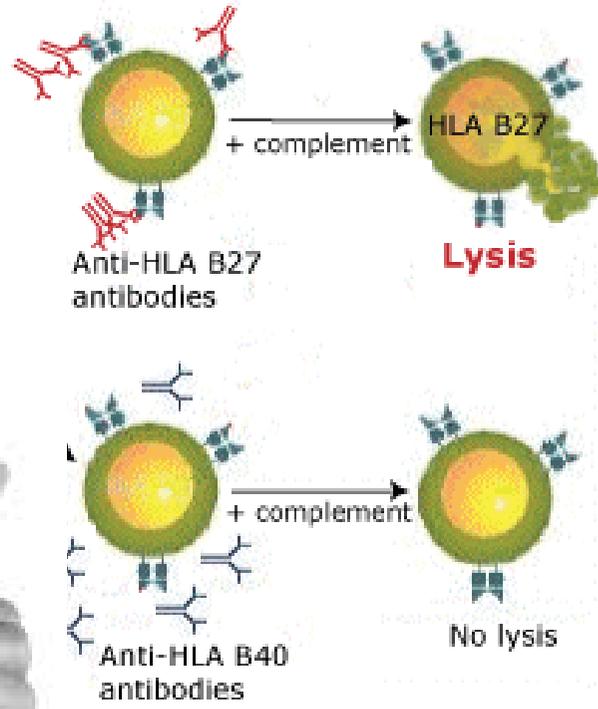
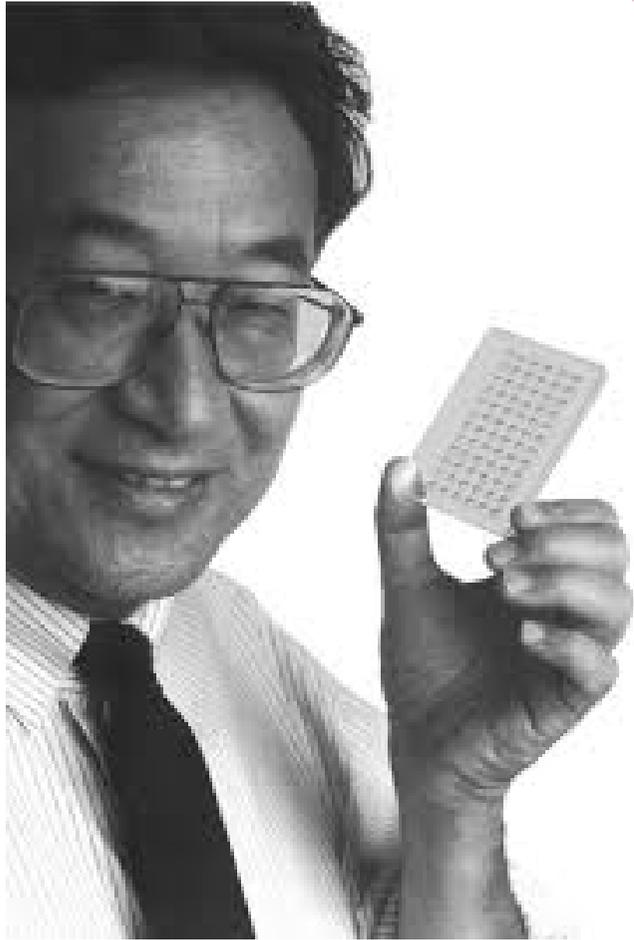
HLA TYPING METHODS

SEROLOGICAL TYPING

Serological Typing

Lymphocytes are **HLA-typed** by crossmatching to panel reactive antibodies (**PRA**) using the complement-dependent cytotoxicity (**CDC**) test.





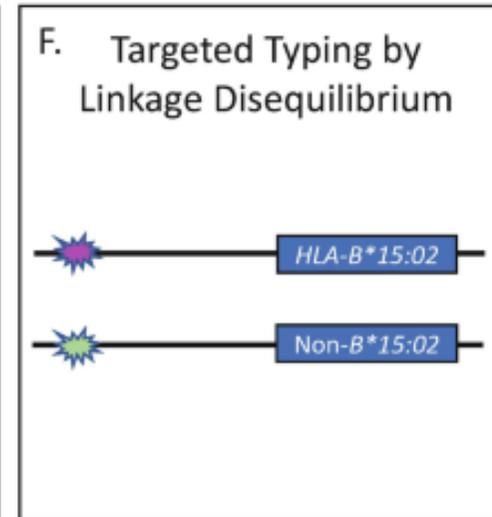
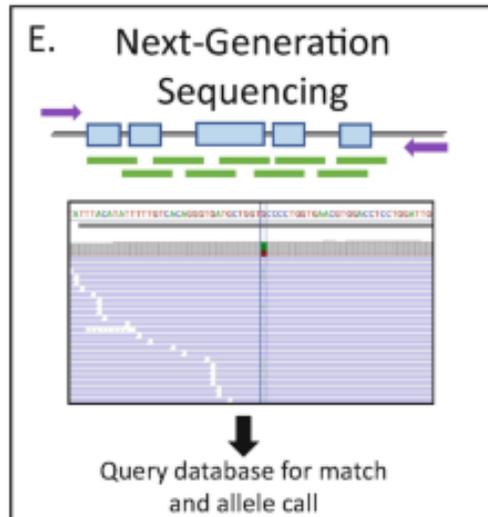
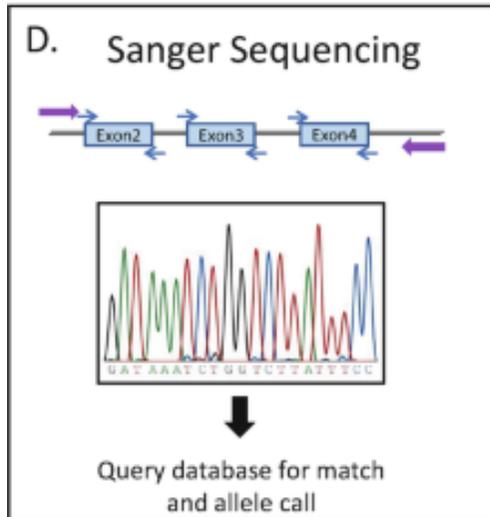
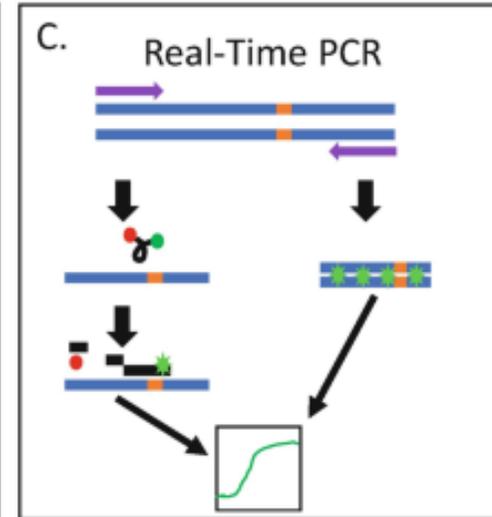
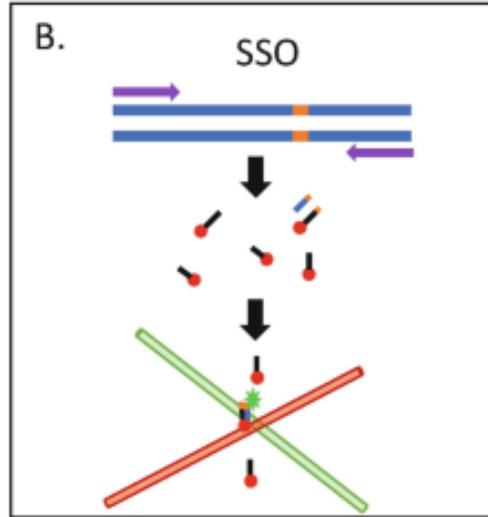
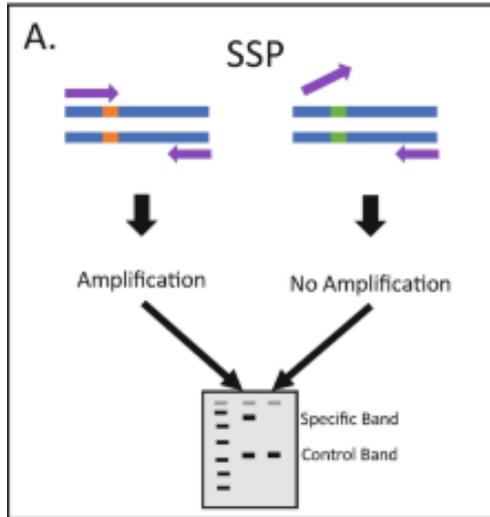
	Pos. Well	Serum Lot	Anti-HLA	Reaktion Reaction	Pos. Well	Serum Lot	Anti-HLA	Reaktion Reaction
	1A	111 H 0150	pos. Control		7A	304 H 0080	A32	
	1B	110 H 0040	neg. Control		7B	304 H 0310	A32+25	
	1C	109 H 0400	A1+36		7C	105 P 0180	A32(25) s.t.wk	
	1D	109 H 0010	A1		7D	303 P 1050 ⁺	A32+74+1+3+11+36 ↓	
	1E	109 H 0630	A1		7E	106 P 0350	A33+10 (28, 11, B63)	
	1F	110 H 0070	A2		7F	110 H 0610	A33+34	
	2F	110 H 0080	A2		8F	106 H 0410	A33+31 (29)	
	2E	105 P 0430	A2+69		8E	204 P 0850	A33	
	2D	306 H 0390	A3		8D	204 P 1700	A34	
	2C	304 H 0250 ⁺	A3		8C	110 H 0430	B 51	
	2B	109 P 0220	A3 (11)		8B	108 H 0380	B 51	
	2A	109 P 0110	A 23		8A	111 H 0130	B5[51+52]	
	3A	109 P 0120	A 23		9A	109 H 0650	B5 [51+52]	
	3B	403 H 1040	A9[23+24]+80		9B	105 H 0540	B5[51+52]+53 (B49, A25)	
	3C	111 H 0190	A 24+2403 s.t.wk		9C	106 H 0580	B7 (81)	
	3D	208 H 0160	A 24+2403		9D	110 H 0130	B7	
	3E	110 P 0210	A 25		9E	105 H 0820	B7+27+81+73+42	
	3F	110 H 0260	A 25		9F	110 H 0280	B8	
	4F	112 P 0200	A10[25+26+34+66]+43		10F	106 H 0550	B8+59	
	4E	112 H 0310	A 25+26+66		10E	106 H 0530	B8+14+39	
	4D	104 H 1340	A 26+wk 66		10D	110 P 0250	B 44	
	4C	105 P 0350	A 26		10C	110 H 0470	B 44	

HLA DICTIONARY

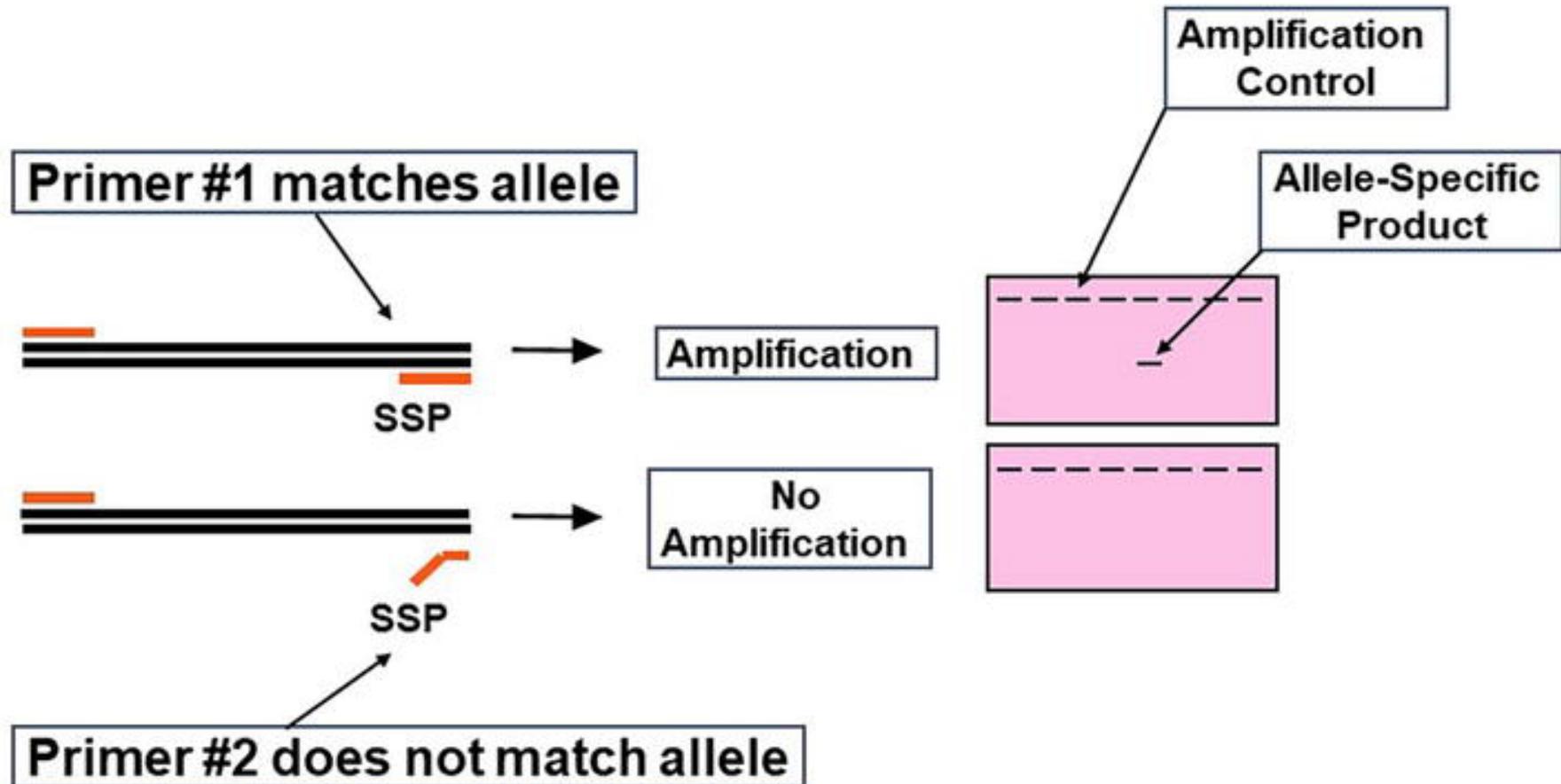
- Serologic : B15 : B62, B63, B75, B76, B77 Molecular: B*15
- Serologic: B70: B71, B72 Molecular: B*15
- Serologic: A28: A68, A69 Molecular: A*68 or A*69
- Serologic: B40: B60, B61 Molecular: B*40

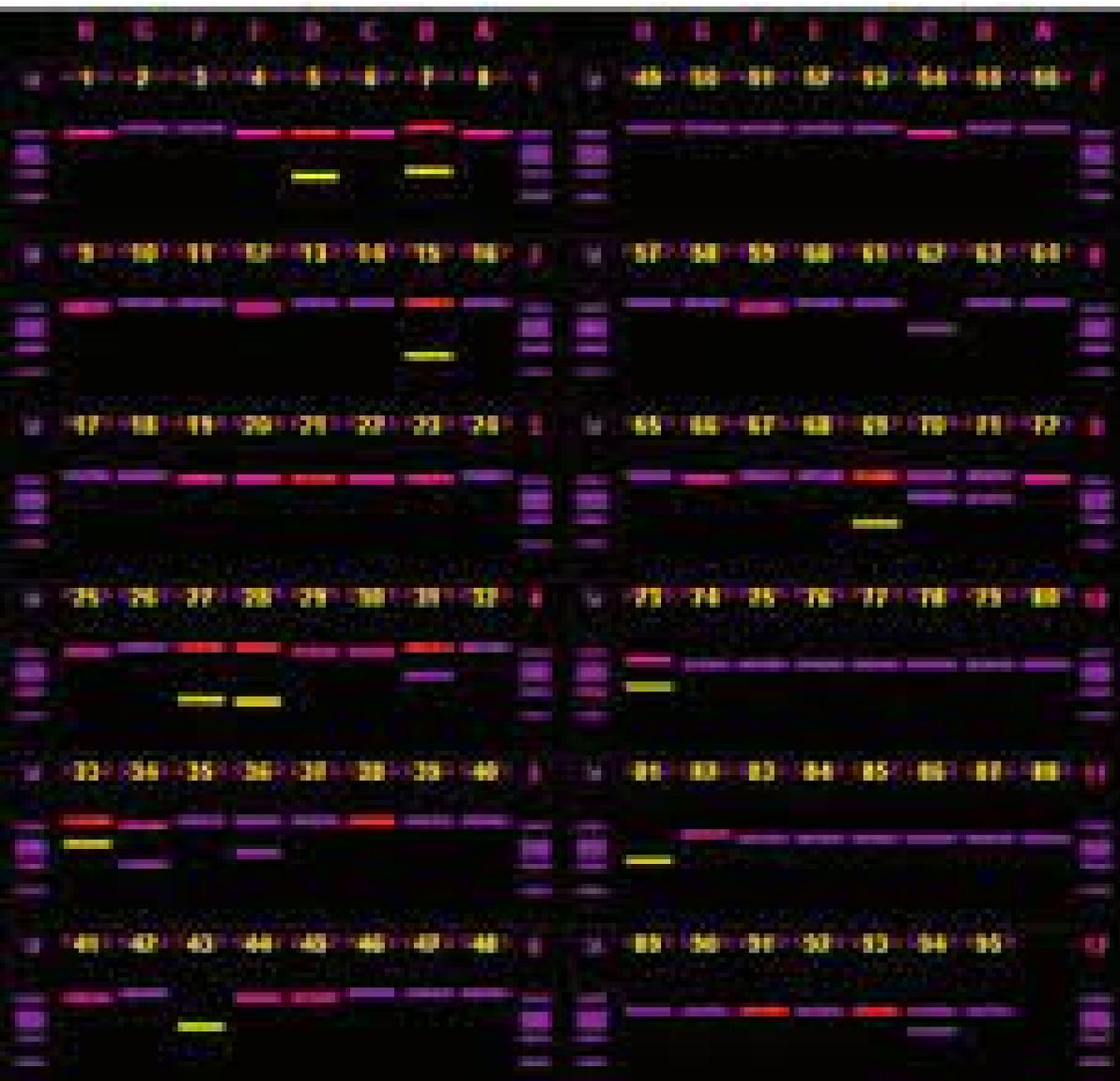
LIMITATIONS

MOLECULAR TYPING METHODS



PCR- SEQUENCE-SPECIFIC PRIMERS (PCR-SSP)





... [Navigation menu: Home, Policies/terms, Out, 88%, Analysis, Typing help, SSP details, Help]

HLAMBPC

SCORE™

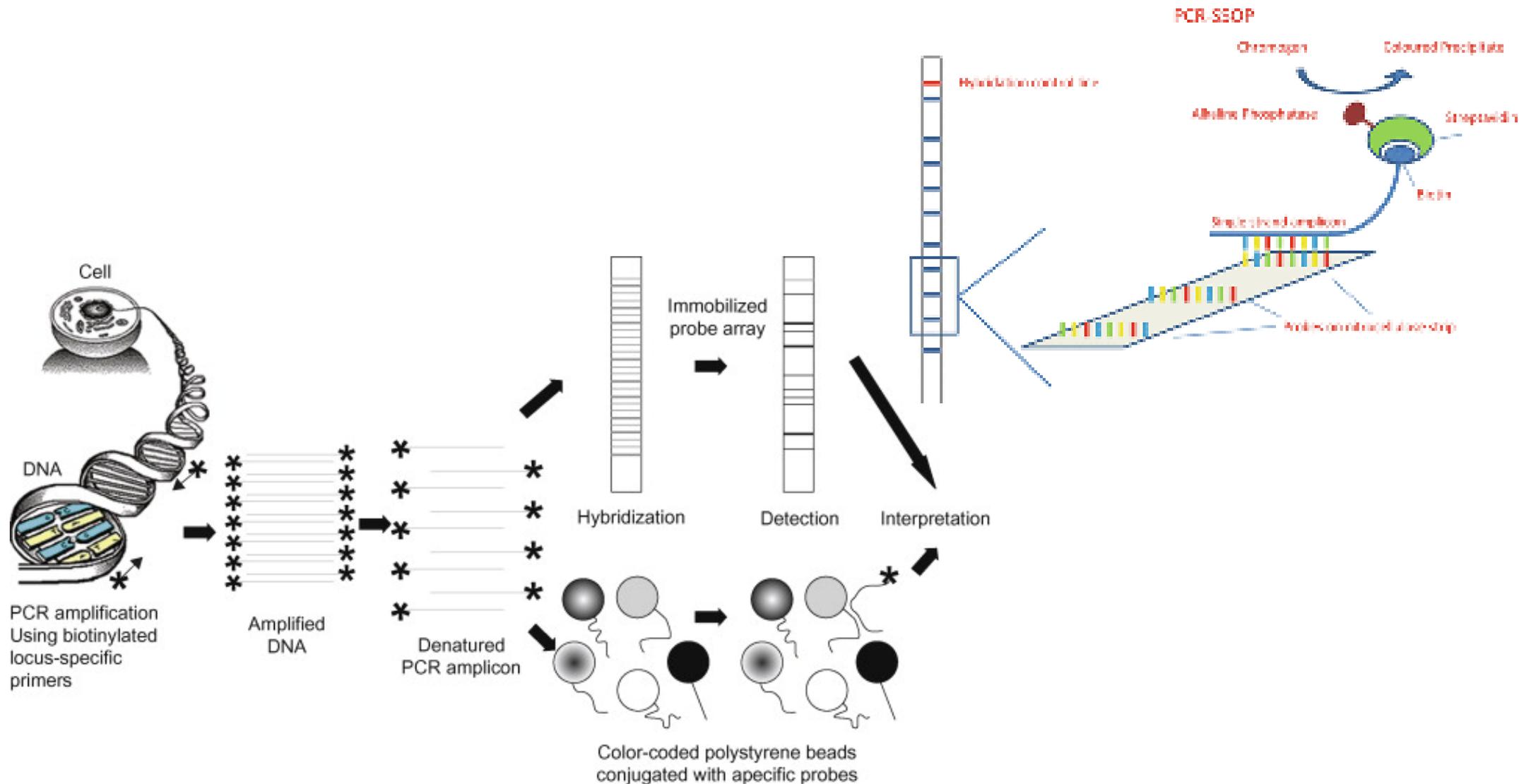
Sequence Completion and Rearrangement Evaluation
for research only

Score:	4.000	<input type="checkbox"/> score_new@COE@vda
User database:	4.001	<input type="checkbox"/> SCORE_pos@vda
Anchor database:	4.001	<input type="checkbox"/> COE@vda
Allele database:	allele@HLAMBPC_2@vda	<input type="checkbox"/> COE@vda

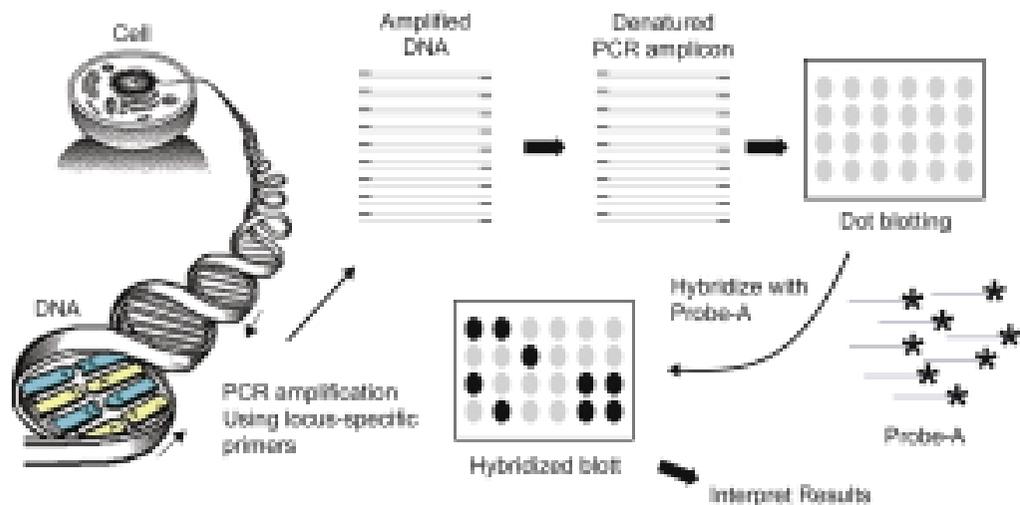
visit the dbMHC resource: <http://www.scorebioinformatics.org/>

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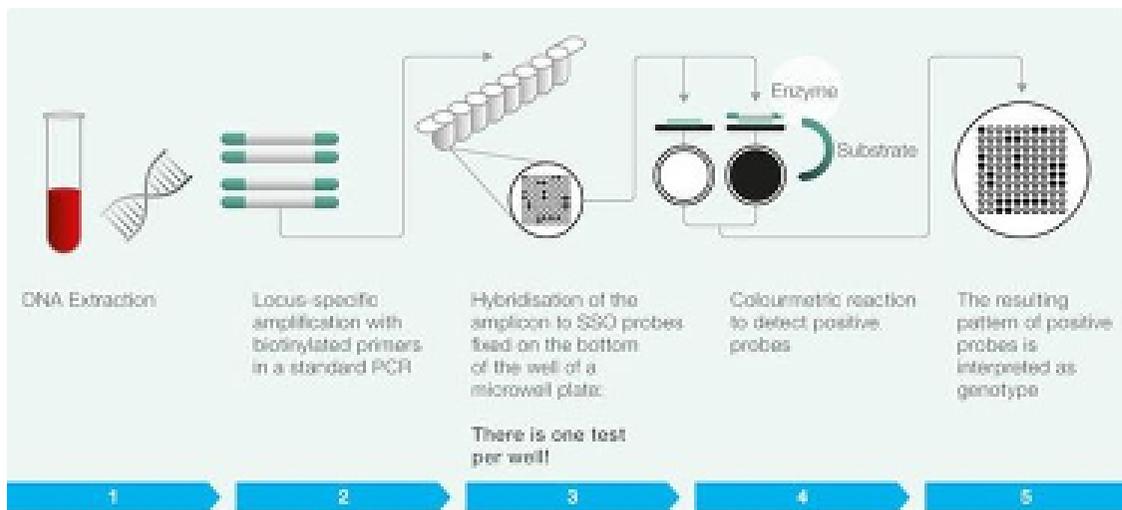
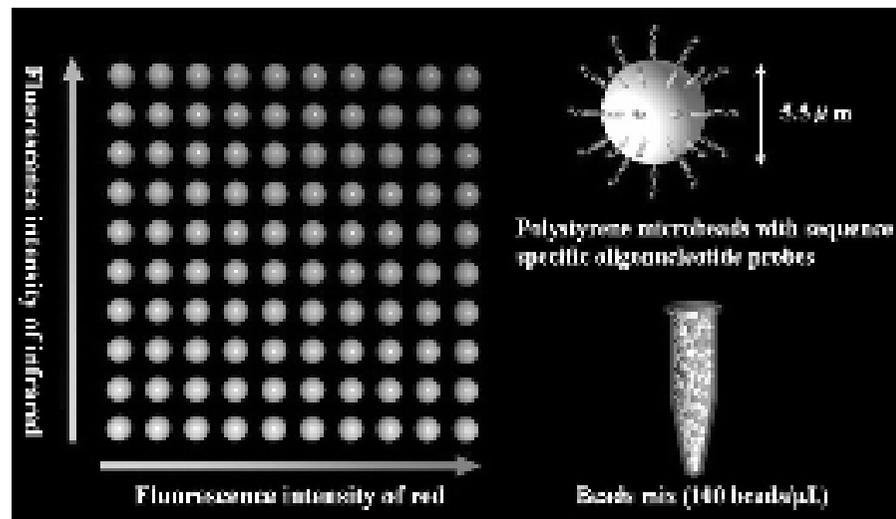
Polymerase chain reaction Sequence-specific Oligonucleotides Probes (PCR-SSOP)



Enzyme-based Protocols



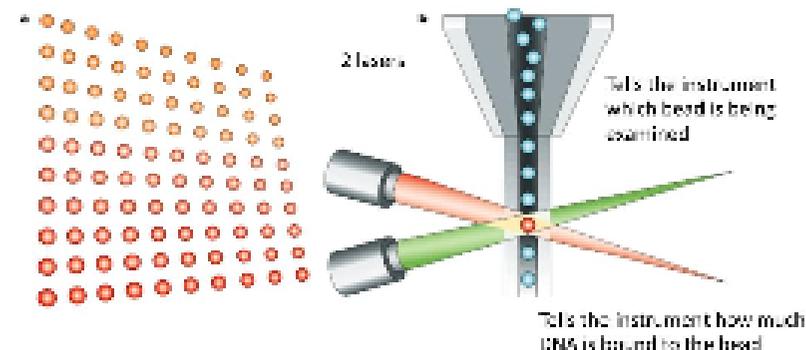
HLA Typing by SSO using Fluorescence



HLA typing by SSO using Luminex platform

100 types of microspheres distinguished by fluorescence emission signature

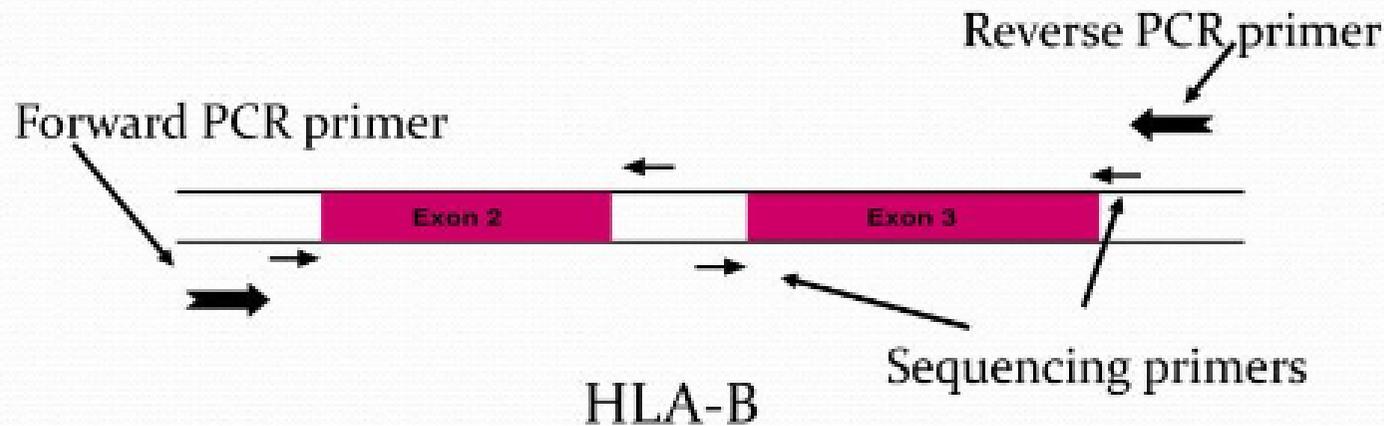
Each microsphere type is coated with different sequence specific oligonucleotide (HLA allele)



Polymerase chain reaction- Sequencing Based Typing (PCR-SBT)

DNA-Based Typing Methods: Sequence-Based Typing

- Sequence-based typing (SBT) is high resolution.
- Polymorphic regions are amplified by PCR and then sequenced.



Next generation sequencing (NGS)

Roche/454 FLX
Pyrosequencing



Illumina



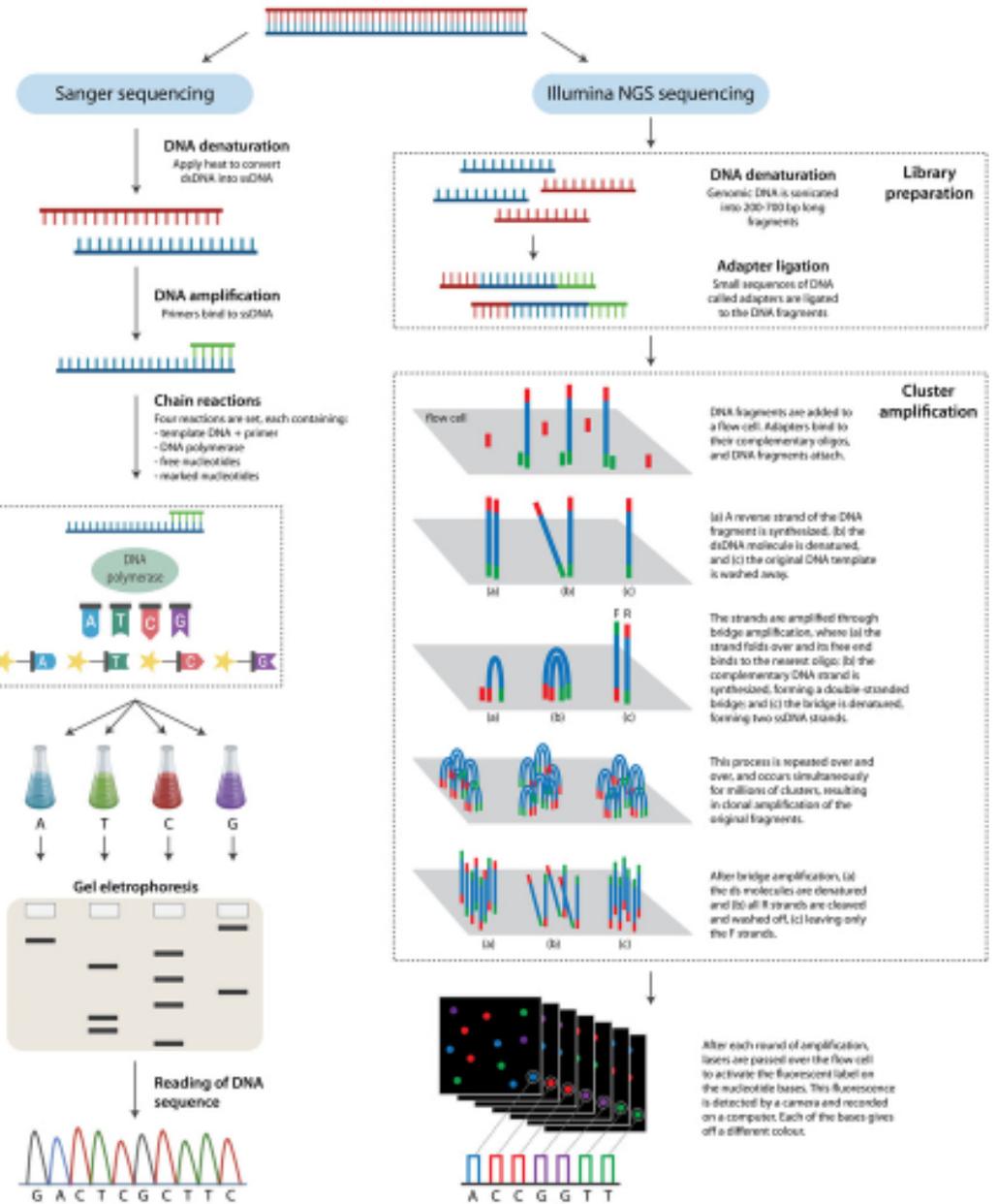
Ion Torrent

ROCHE/454 FLX PYROSEQUENCING

- The GS FLX system utilizes forward and reverse PCR primers fused to adaptors that enable the capture of a single-stranded DNA to a bead in the well of a picotiter plate (PTP).
- The PTP has 1.6×10^6 wells, one bead per well where water-in-oil emulsion PCR occurs.
- After PCR, each well will contain millions of copies of the same DNA strand.
- A number of enzymes (polymerase, luciferase, sulphurylase and apyrase) are added to the wells by the GS FLX system followed by sequential addition of nucleotides.
- Incorporation of one or more nucleotides results in the release of pyrophosphate, which is converted to ATP by sulphurylase. ATP is used by luciferase to create light.
- The sequential addition of nucleotides and the release of light are monitored by a CCD camera, and a flowgram is produced, similar to a sequencing chromatogram, showing the order of A, C, G, T.
- Through clonal PCR and massively parallel pyrosequencing, unambiguous HLA typing is achievable.

ION TORRENT

- Ion Torrent's Personal Genome Machine™ detects H⁺ released upon the sequential addition of (unlabelled, natural) nucleotides.
- Release of a proton causes a change in pH, producing an electrical signal that can be measured.
- This technology therefore does not require any optical detection system or enzymatic amplification cascades and thus eliminates sources of sequencing errors while delivering highly uniform genome coverage in a very short time (2h)



A new generation of molecular sensing technology

Explore the nanopore portfolio of flow cells, compatible devices, manual and automated library prep, and data analysis solutions.



PREPARE your library

[Explore >](#)



SEQUENCE with flow cells & devices

[Explore >](#)



ANALYSE your data

[Explore >](#)





MinION

Size: W 105, H 23, D 33 mm
Weight: 87 g

(A)



SmidgION

(B)



GridION

Size: W 365 H 220 D 360 mm
Weight: 11 kg

(C)



PromethION

Size: W 440, H 400, D 240 mm
Weight: 40 kg

(D)



Array of microcaffolds

Sensor chip

ASIC

Flow cell

1

A MinION flow cell contains 2,048 nanopores used to sequence DNA or RNA. The wells are inserted into an electrically resistant polymer membrane supported by an array of microcaffolds connected to a sensor chip. Each channel associates with a separate electrode in the sensor chip and is controlled and measured individually by the application-specific integration circuit (ASIC).

2

Ionic current passes through the nanopore because a constant voltage is applied across the membrane, where the trans side is positively charged.



trans

ions

C G A T

double-stranded DNA (dsDNA)

Motor protein

Nanopore

3

Under the control of a motor protein, a double-stranded DNA (dsDNA) molecule (or an RNA-DNA hybrid duplex) is first unwound.



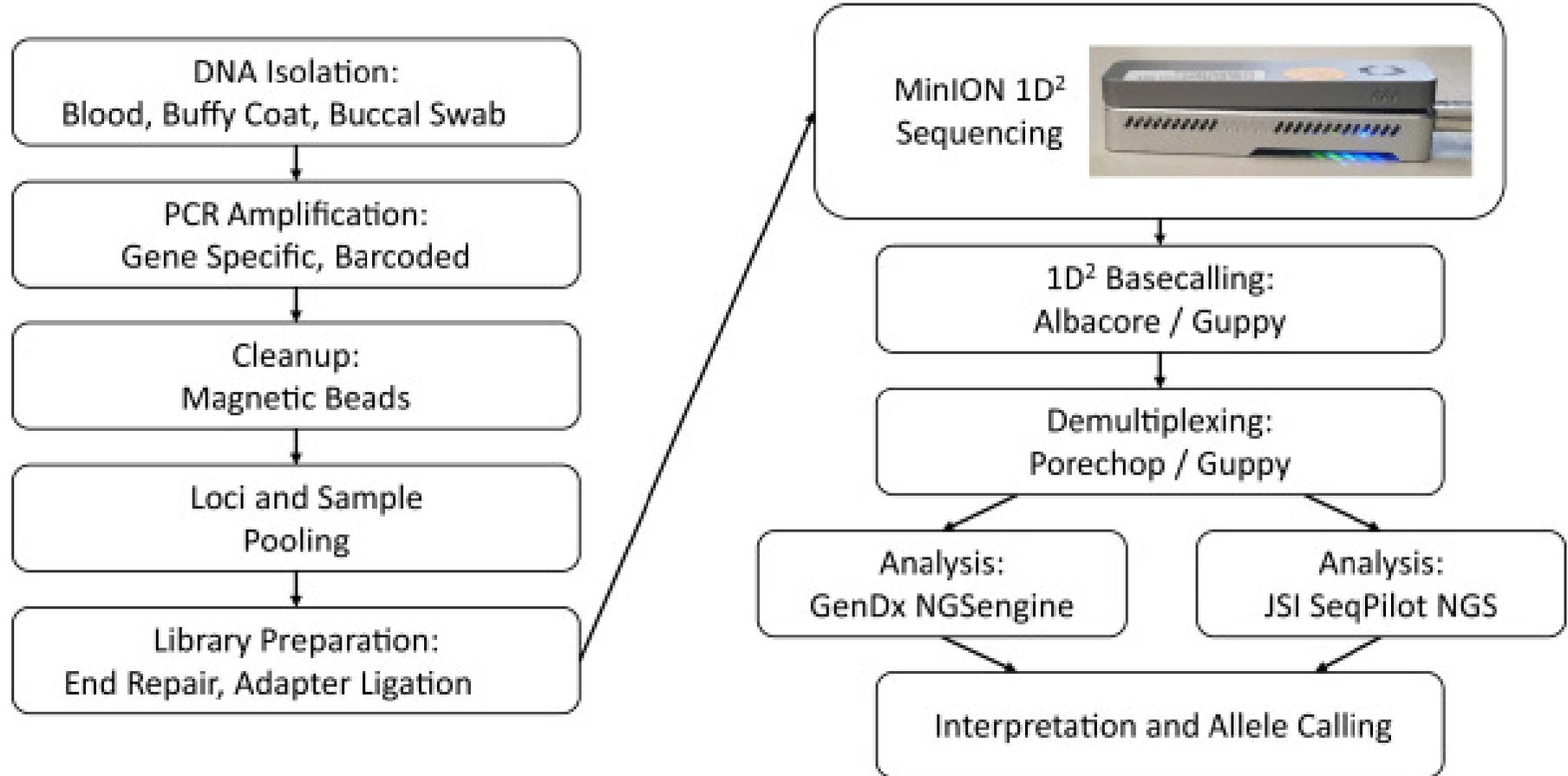
4

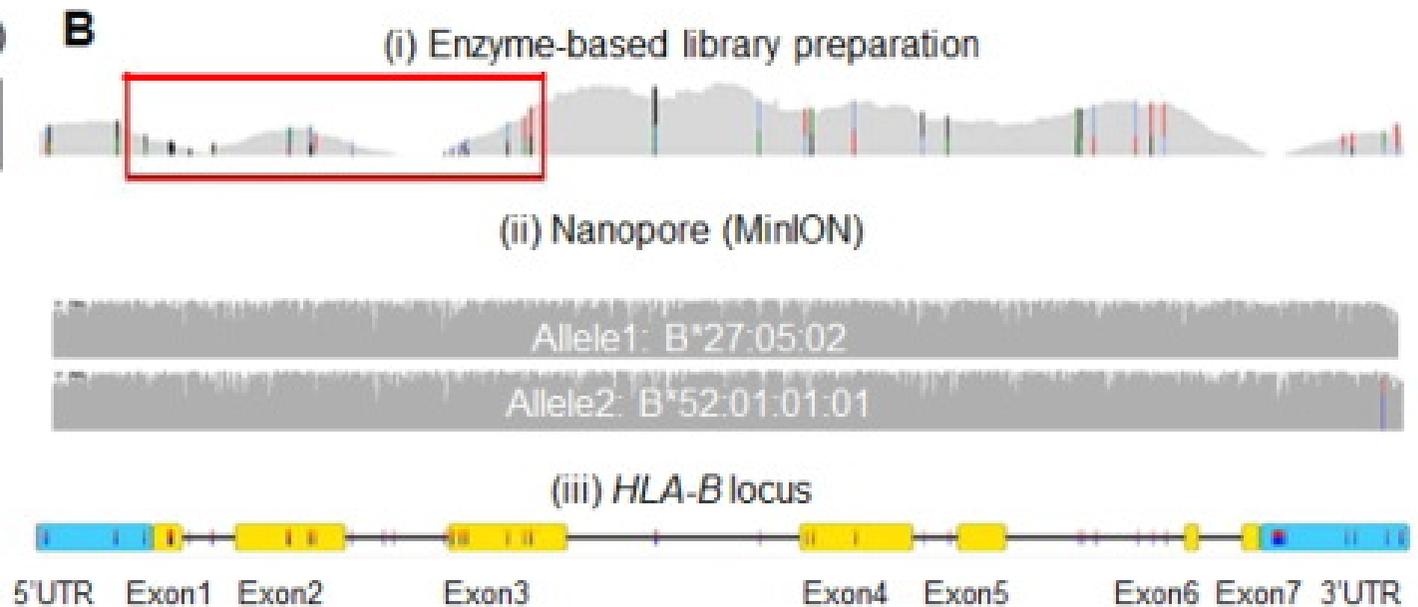
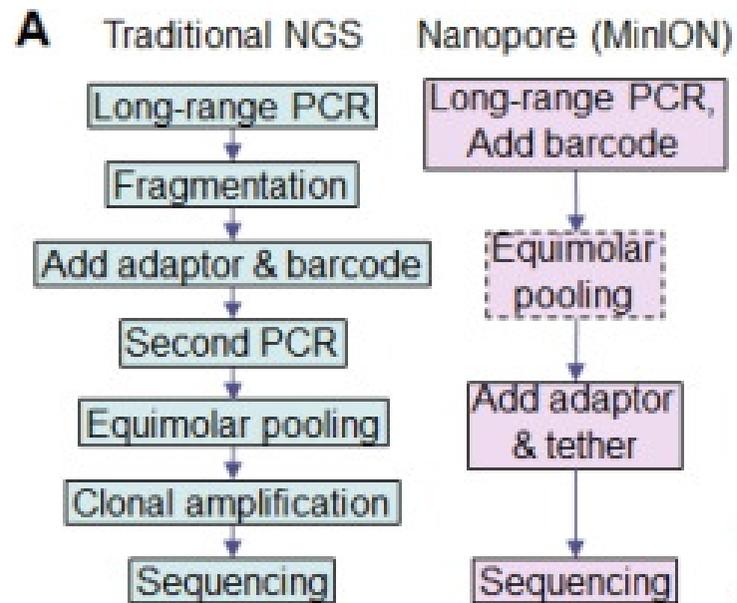
Then a single-stranded DNA with a negative charge is ratcheted through the nanopore, driven by the voltage. As nucleotides pass through the nanopore, a characteristic current change is measured and is used to determine the corresponding nucleotide sequence.

MinION

OXFORD NANOPORE TECHNOLOGY

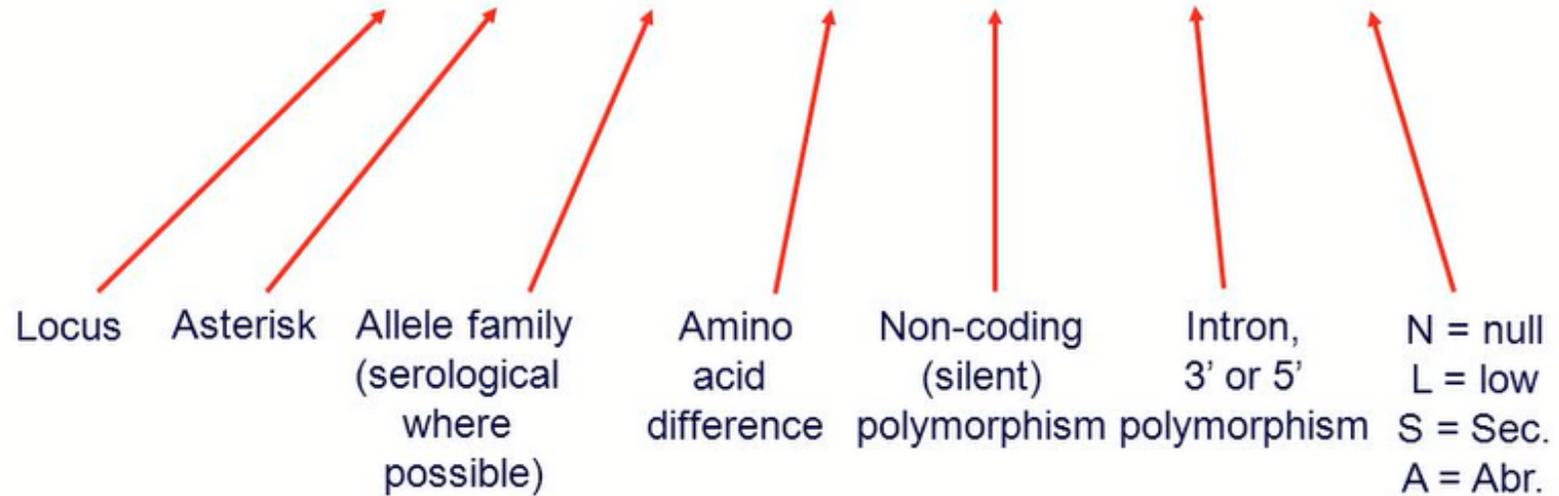
- Oxford Nanopore Technologies (Oxford, UK) utilize an engineered protein nanopore, alpha-haemolysin, adapted so that the identity of DNA bases can be ascertained as they pass through the nanopore by measuring variations in an ionic current passing through the pore.
- The company is developing two methods – ‘exonuclease’ sequencing where individual bases are cleaved by the enzyme and passed through the nanopore in sequence and ‘strand’ sequencing where an ssDNA molecule is ratcheted through the nanopore by the enzyme





HLA Allele Nomenclature

HLA - A * 24 02 01 01
HLA - A * 24 02 01 02 L

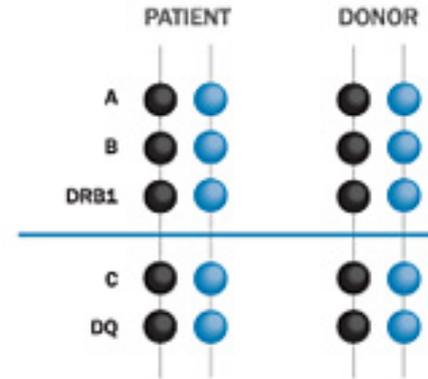


HLA MATCHING

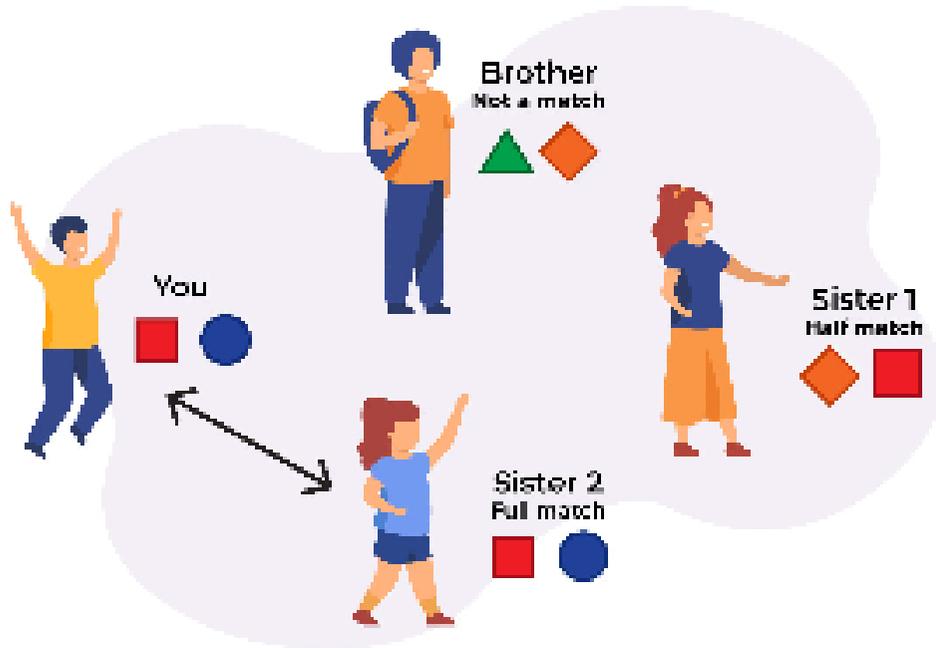
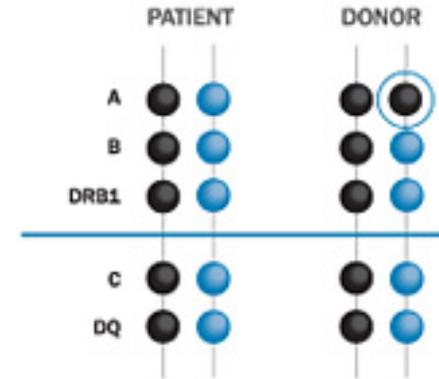
46

BEST MATCHED DONOR

A. 6 of 6 Match / 10 of 10 Match

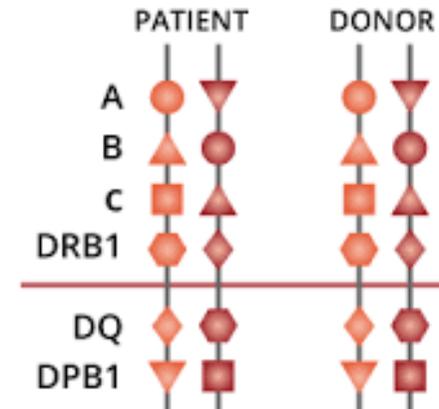


B. 5 of 6 Match / 9 of 10 Match

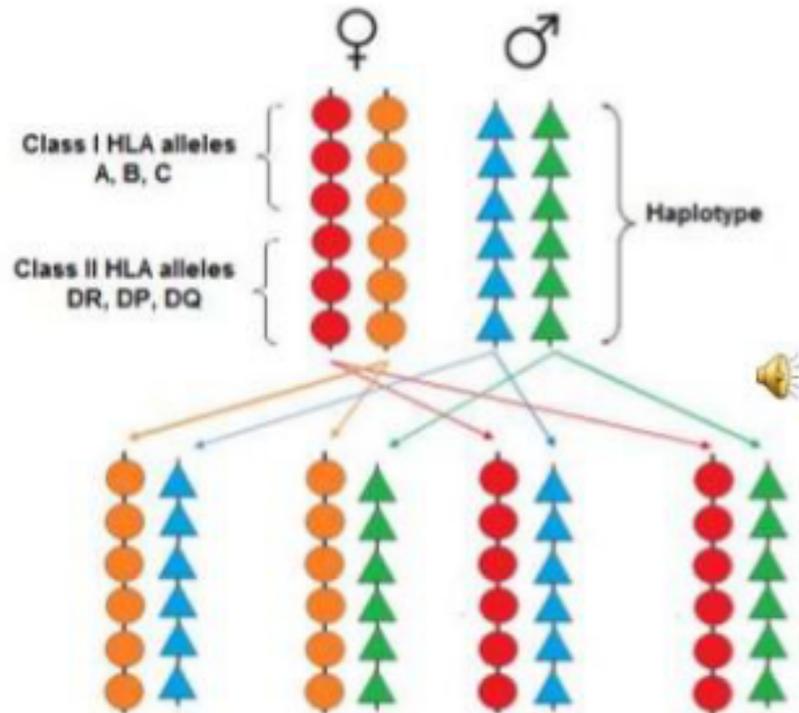


Related

12 / 12 Match



HLA inheritance and familial matching

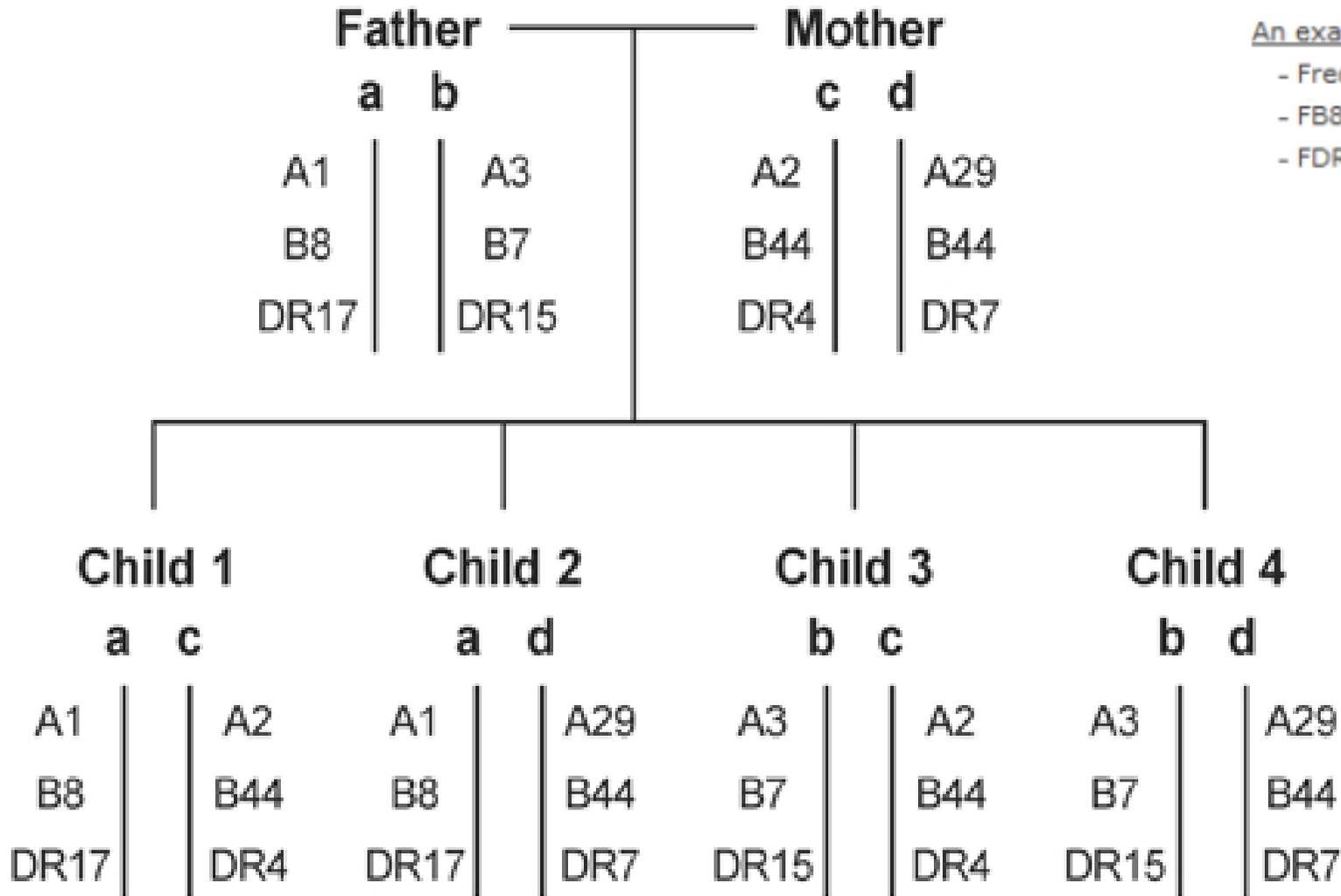


- 'Haplotype': a combination of alleles at adjacent loci on a chromosome that are inherited together.

- Breeding between two heterozygous individuals results in children with 4 possible genotypes

- There is a 1 in 4 chance of a complete match between two siblings

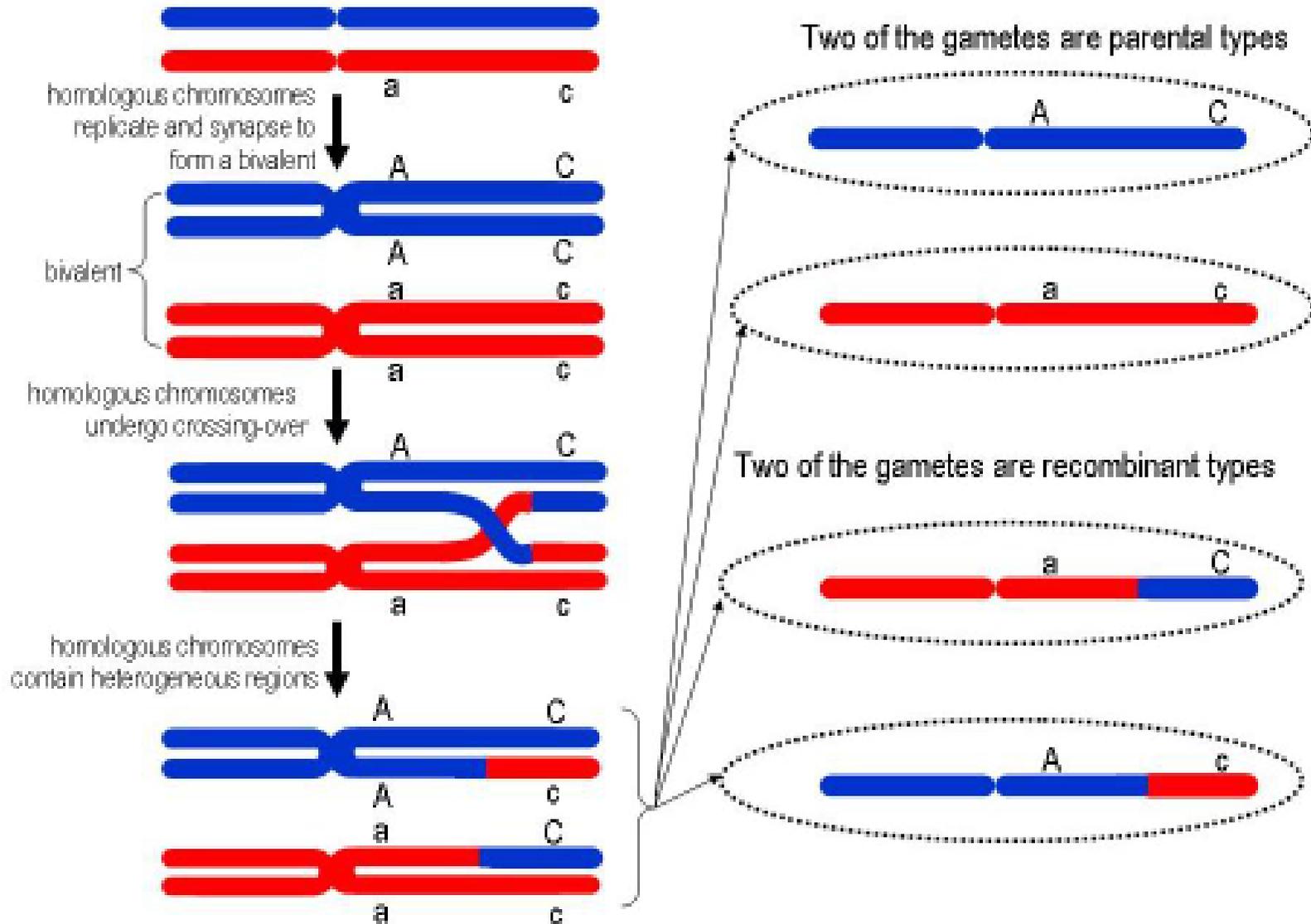
Linkage Disequilibrium



An example: Haplotype A1-B8-DR3

- Frequency of A1 (FA1) = 0.137
- FB8 = 0.103
- FDR3 = 0.11
- Expected of A1-B8-DR3 = **0.0015**
- Observed = **0.025**

RECOMBINATION (CROSSING OVER)



+ Add image

Classic HLA loci

High expression loci (HEL)

- **A**
- **B**
- **C**
- **DRB1**

Low expression loci (LEL)

- DRB3
- DRB4
- DRB5
- DQA1
- **DQB1**
- DDPA1
- **DPB1**

Matched sibling donors

- ▶ **Primary low resolution typing for all available first degree relatives.**
- ▶ **Low or high resolution confirmatory typing of the patient and phenotypically matched donor based on HLA pedigree.**
- ▶ **In most cases high resolution typing is not needed.**

Matched other related donors

- ▶ Typing of second degree relatives or beyond based on intra familial marriages.
- ▶ Low or high resolution confirmatory typing of the patient and phenotypically matched donor based on HLA pedigree.
- ▶ In most cases high resolution typing is needed.

Results

- Overall, outcomes for patients treated from a 1-antigen/allele mismatch related donor were significantly worse than from a MUD, primarily because of increased NRM.
- Overall survival (OS) rates at 3 years for 1-antigen/allele mismatched related donor and MUD transplant recipients were 19% and 45% ($P = .007$), and NRM rates were 40% and 26% ($P = .05$), respectively.
- Patients with class I mismatches appeared to have poorer OS than did patients with class II mismatches.
- A higher incidence of graft rejection was identified in the mismatched related donor group ($P = .02$).

An Apparently 8/8 But Really Haploidentical Donor

► Patient and his uncle:

A*02,*24; B*35,*51; C*04,*16; DRB1*11,*12

► High resolution HLA typing results:

► Patient: A*24:02:01 B*35:01 DRB1*11:04

► Uncle: A*24:02:01 B*35:02 DRB1*11:01

Split HLA Antigens

- A9: A23, A24
- A10: A25, A26, A34, A66
- A19: A29, A30, A31, A32, A33, A74
- A28: A68, A69
- B5: B51, B52
- B12: B44, B45
- B14: B64, B65
- B15: B62, B63, B75, B76, B77
- B16: B38, B39
- B17: B57, B58
- B21: B49, B50
- B22: B54, B55, B56
- B40: B60, B61
- B70: B71, B72

Linkage Disequilibrium/ Haplotypes

- LD: Alleles occur together with a greater frequency than would be expected by chance
 - B/C strong, DR/DQ strong, A less strong, DP weak
 - e.g. B*0801 - 99% will be Cw*0701
 - But B*1801 either Cw*0501, *0701
 - e.g DRB1*1501 - will be DQB1*0602
 - But DRB1*0401 either Cw*0301, *0302
- Haplotype: A group of genes inherited together
 - e.g. A*0101,B*0801,Cw*0701,DRB1*0301,DQB1*0201

Example

- Patient:
 - A*0201, B*1801, DRB1*0401
- Finding a donor:
 - Less strong LD - therefore ‘NOT predictable’
i.e. equal chance of C being *0701, *0501,
*1203; DQB1 50/50 chance of *0301, *0302
 - At low resolution (A2, B18, DRB4) unable to
predict this will match

G CODES FOR REPORTING OF AMBIGUOUS ALLELE TYPING

- HLA alleles that have identical nucleotide sequences across the exons encoding the peptide binding domains (exon 2 and 3 for HLA class I and exon 2 only for HLA class II alleles), will be designated by an upper case 'G' which follows the first 3 fields of the allele designation of the lowest numbered allele in the group.
- The group designation will contain a minimum of six digits.

HLA-A		HLA-B		HLA-C		HLA-DRB1		HLA-DRB345		HLA-DQB1		HLA-DPB1	
A*01:04N*	WD	B*07:67N	WD	C*02:38N	WD	DRB1*07:10N	WD	DRB4*01:03:01:02N*	WD	DQB1*02:18N	WD	DPB1*61:01N	WD
A*01:15N	WD	B*07:181N	WD	C*02:92N	WD	DRB1*07:26N	WD	DRB4*01:16N	WD	DQB1*02:20N	WD	DPB1*64:01N	WD
A*01:16N	WD	B*14:07N	WD	C*04:09N*	I	DRB1*12:24N	WD	DRB4*02:01N	WD	DQB1*03:118N	WD	DPB1*120:01N	WD
A*01:57N	WD	B*15:01:01:02N*	WD	C*04:93N	WD			DRB4*03:01N	WD	DQB1*06:26N	WD	DPB1*154:01N	WD
A*01:123N	WD	B*15:79N	WD	C*04:95N	WD			DRB5*01:08N*	WD	DQB1*06:75N	WD	DPB1*161:01N	WD
A*02:53N	WD	B*15:181N	WD	C*05:07N	I			DRB5*01:10N	WD	DQB1*06:77N	WD	DPB1*218:01N	WD
A*02:83N*	WD	B*15:190N	WD	C*05:99N	WD					DQB1*06:144N	WD	DPB1*357:01N	WD
A*02:94N	WD	B*35:165N	WD	C*06:16N	WD							DPB1*570:01N	WD
A*02:113N	WD	B*37:03N	WD	C*06:79N	WD								
A*02:125N	WD	B*37:42N	WD	C*07:32N	WD								
A*02:227N	WD	B*39:40N	WD	C*07:33N	WD								
A*02:514N	WD	B*40:22N	WD	C*07:55N	WD								
A*03:21N*	WD	B*40:142N	WD	C*07:61N	WD								
A*11:21N*	WD	B*40:155N*	WD	C*07:104N	WD								
A*11:109N	WD	B*44:23N	WD	C*07:198N	WD								
A*23:11N	WD	B*51:11N*	WD	C*07:227N	WD								
A*23:19N	WD			C*07:452N	WD								
A*24:09N*	WD			C*08:127N	WD								
A*24:11N*	WD			C*15:122N	WD								
A*24:36N	WD			C*16:30N	WD								
A*24:84N	WD												
A*24:90N	WD												
A*24:252N	WD												
A*25:12N	WD												
A*30:70N	WD												
A*30:78N	WD												
A*31:14N*	WD												
A*31:60N	WD												
A*32:27N	WD												
A*32:45N	WD												
A*34:10N	WD												
A*68:18N	WD												

HLA disease association

Panel #	Disease or Hypersensitivity	Page #	Associated HLA Allele
1	Abacavir Hypersensitivity	3	HLA B*57:01
2	Allopurinol Hypersensitivity	4	HLA B*58:01
3	Spondyloarthropathies (Ankylosing Spondylitis, Reactive Arthritis, Juvenile Arthritis)	7	HLA-B27
4	Eye Disease HLA Association (Acute Anterior Uveitis HLA-B27, Behçet's Disease HLA-B51, Birdshot chorioretinopathy HLA-A29)	11	HLA-B27 HLA-B51, HLA-A29
5	Carbamazepine Hypersensitivity	16	HLA-B*15:02, HLA-A*31:01
6	Celiac Disease (Celiac Genetics)	18	HLA-DQA1 and HLA-DQB1 associated alleles and DQ2/DQ8 heterodimers
7	Narcolepsy	24	HLA-DQB1*06:02, DQA1*01:02
8	Pediatric Acute-onset Neuropsychiatric Syndrome (PANS)	27	HLA-B38, HLA-B5, HLA-B51, HLA-B52, HLA B27, HLA-Bw4
9	Psoriasis	29	HLA-B27, HLA-Cw6
10	IL-1 and IL-6 inhibitors Hypersensitivity	32	HLA-DRB1*15:01, HLA-DRB5*01:01 HLA-DQB1*06:02, HLA-DQA1*01:02,

مرکز پذیره نویسی اهداکنندگان سلولهای بنیادی و بانک HLA

مرکز تحقیقات ایمنولوژی، آسم و آلرژی

11. تهران، مرکز پذیره نویسی اهداکنندگان سلولهای بنیادی خونساز مرکز تحقیقات ایمونولوژی، آسم و آلرژی، مرکز طبی کودکان دانشگاه علوم پزشکی تهران

تهران، انتهای بلوار کشاورز خیابان دکترقرب، بیمارستان مرکز طبی کودکان، ساختمان شماره3، طبقه 4.

۰۲۱-۶۶۹۳۵۸۵۵۰۲۱۶۶۹۰۷۴۱۵

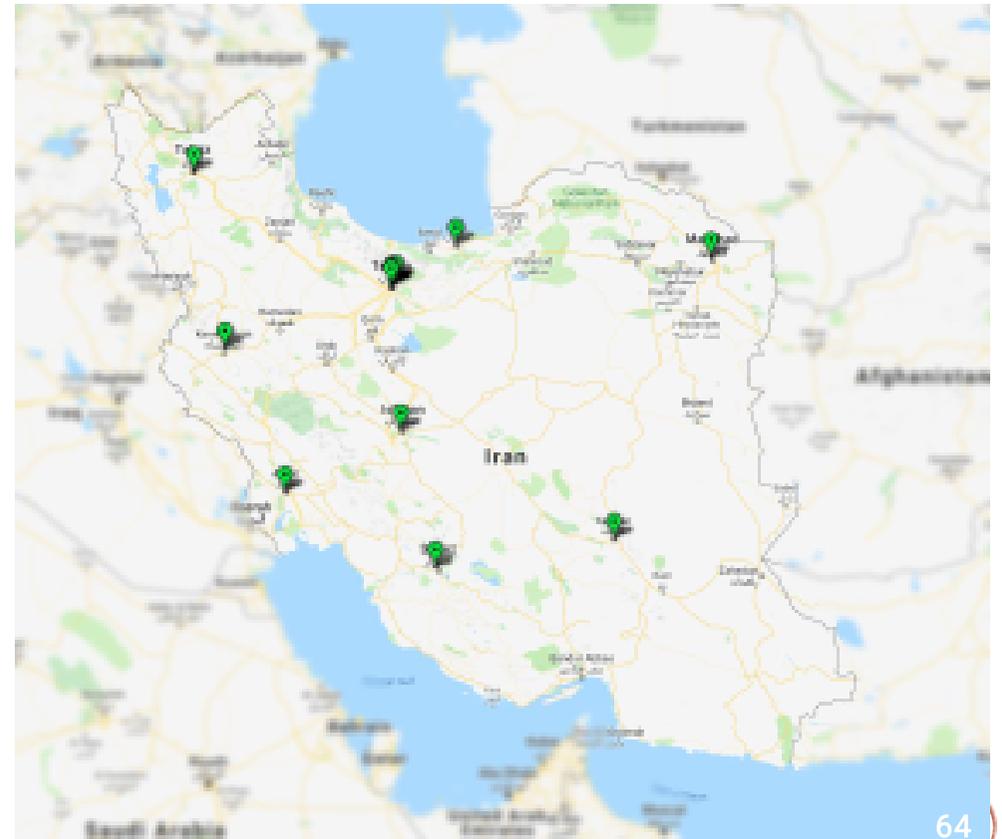
جمهوری اسلامی ایران
دانشگاه علوم پزشکی تهران

شبکه ملی اهداکنندگان سلول های بنیادی خونساز ایران
Iranian National Stem Cell Donor Network

صفحه اصلی | درباره ما | گالری | متخصصین | اهداکنندگان | بیماران | حامیان و خیرین

بنی آدم اعضای یکدیگرند

بانک های خون بند ناف | مراکز پیوند سلول های بنیادی خونساز | مراکز پذیره نویسی



درخواستها

درخواست جستجو

لیست پذیرش

پذیرش

صفحه اصلی

لیست پذیرش

Page 1 of 58 (2580 items)

ردیف	نام خانوادگی	نام	نام پدر	تاریخ تولد	کد ملی
001	نوروزی زاده	فریم	Hanah		
002	موسویان	ساجده		1397/1/21	
003	طاهری	پروین			
004	احمدی	اوان			
005	صالحی	فاطمه		1397/1/21	
006	ملکی	آناهید		1397/1/21	
007	نار آقایی	مهیار		1397/1/21	
008	محمد نور شهبازی	علیا		1397/1/21	
009	شهباز	فریبا			
010	کفایه شهبازی	شیدا			
011	احمد نور شهبازی	محمد رضا		1397/1/21	
012	داجک	شاهین		1397/1/21	
013	سبی زاده مرشدی	فاطمه		1397/1/21	
014	مختار شهبازی	شیرین		1397/1/21	
015	شکوفه شهبازی	راجه		1397/1/21	
016	طاهری	وردانه		1397/1/21	
017	ساجدیان	فریم			
018	اکبری	فریم		1397/1/21	



جمهوری اسلامی ایران
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شبکه ملی اهداکنندگان سلول های بنیادی خونساز ایران

Iranian National Stem Cell Donor Network



حامیان و خیرین

بیماران

اهداکنندگان

متخصصین

گالری

درباره ما

صفحه اصلی



از مهر شما سپاسگزاریم...

~~۶۳,۸۲۴~~

حدود 80 هزار نفر

اهداکننده بزرگسال و خون بند ناف

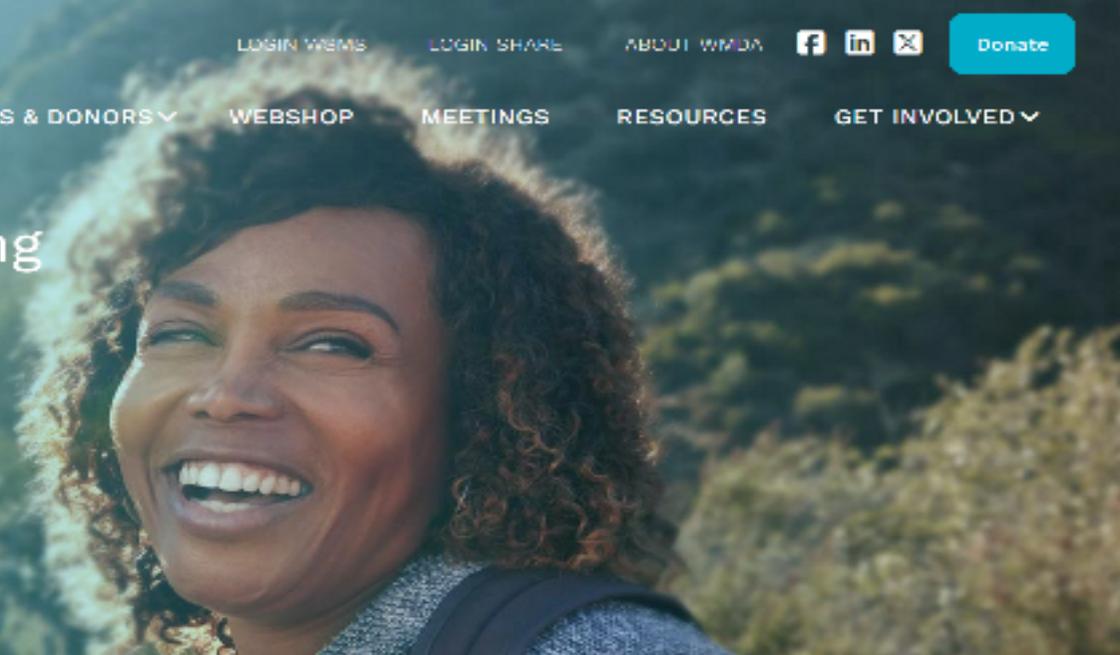
تا سال ۱۳۹۸



The Global Database Listing over 41 million potential cures

4 1 . 8 2 1 . 7 9 4

AMOUNT OF CURRENT DONORS AND CORD BLOOD UNITS



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Donor registration


German donor centers


Current
donor numbers:
10.254.730


Support us


ZKRD for
professionals

مشخصات درخواست

نام و نام خانوادگی	نام پدر	نام و نام خانوادگی	نام پدر
نام و نام خانوادگی	نام پدر	نام و نام خانوادگی	نام پدر

مشخصات بیمار

نام خانوادگی	نام پدر	نام خانوادگی	نام پدر
نام خانوادگی	نام پدر	نام خانوادگی	نام پدر
نام خانوادگی	نام پدر	نام خانوادگی	نام پدر
نام خانوادگی	نام پدر	نام خانوادگی	نام پدر
نام خانوادگی	نام پدر	نام خانوادگی	نام پدر
نام خانوادگی	نام پدر	نام خانوادگی	نام پدر

مشخصات بیماری

نام بیماری	شرح بیماری	شرح بیماری	شرح بیماری
نام بیماری	شرح بیماری	شرح بیماری	شرح بیماری

آزمون سلجش HLA بیمار

First antigen	Second antigen
A	A
B	B
C	C
DRB1	DRB1
DQB1	DQB1
DQA1	DQA1

ارسال



پیام جدید



صفحه اصلی

مراکز

واحد

آزمایش خون

صندوق پیام

بخش بیمار

از طرف

آزمایش اسم و

مرکز تخصصی

دانشگاه



Search Report

Search ID: 20189
Search Date: 1402/12/14 12:56:33
Type: ALL

Patient / Donor	A	B	C	DRB1	DQB1	DPB1
Sex Age BG Rh CMV Status CCR5	01:01:01 02:05:01	14:02:01 15:17:01	07:01:02 08:02:01	01:02:01 13:03:01	01:01:01 05:05:01	04:01:01 47:01
10/10 Match Grade						
1 INSC002000125109 M 34	01 02	14 15		01 10		
9/10 Match Grade						
2 INSC002000112980 M 30	01 31	14 15		01 10		
3 INSC005000074293 M 30	01 02	14 15		01 15		
4 INSC002000154501 M 31	01 02	14 19		01 10		
5 INSC005000133430 M 32	01 32	14 15		01 13		
6 INSC005000099190 F 31	01:XX 31:XX	14:XX 15:XX		01:XX 10:XX		
7 INSC002000154951 M 33	01 6h	14 15		01 10		
8 INSC002000154273 M 35	01 32	14 15		01 13		
9 INSC002000151555 M 37	01 31	14 15		01 10		
10 INSC002000150920 M 37	01 02	14 15		01 11		
11 INSC002000124501 M 38	02 01	14 15		01 10		
12 INSC005000112100 M 39	01:XX 32:XX	14:XX 15:XX		01:XX 13:XX		

مراحل بعدی:

ارسال درخواست پیگیری اهدا کننده سازگار از وزارت بهداشت

تماس با اهدا کننده (گان) سازگار ثبت شده در مرکز پذیره نویسی مرکز IAARI برای
مراجعه ایشان به منظور انجام تستهای تاییدی و تکمیلی (نیاز به پیگیری های مکرر و
برنامه ریزی دقیق دارد، خصوصا در زمانیکه بایستی تست کراس مچ انجام شود)

پیگیری نامه نگاری ها با بخش پیوند و بیمارستان

پیگیری مسایل مالی و دریافت ارز دولتی و راهنمایی بیماران در این خصوص (فعلا ارز
دولتی لغو شده است)

پیگیری وضعیت بیماران

محدودیت ها و موانع پیش رو در مرکز پذیره نویسی:

- ❖ عدم اطلاع رسانی کافی در مورد اهدا و پیوند سلولهای بنیادی به ویژه در مناطق بدون مراکز پذیره نویسی
- ❖ عدم تامین منابع مالی به منظور تهیه کیت و مواد لازم برای انجام HLA Typing برای اهداکننده گان
- ❖ گران بودن تست HLA Typing به ویژه نوع High Resolution که بیمه به آن تعلق نمیگیرد
- ❖ عدم حمایت نهادهای دولتی به ویژه وزارت بهداشت از بیماران در قبال تهیه کیت HLA Typing High Resolution با نرخ دولتی
- ❖ عدم تامین ارز دولتی کافی به منظور سلولگیری و انتقال آن از اهداکنندگان خارجی
- ❖ به روز نبودن اطلاعات مربوط به اهداکنندگان داخلی در سایت مربوط به وزارت بهداشت
- ❖ عدم برنامه ریزی درست در مورد ساماندهی مراکز پذیره نویسی به منظور استفاده مراکز از سیستم اتوماسیون مراکز درمانی در جهت هماهنگی های بیشتر بین بخش های درگیر در پیوند
- ❖ عدم امکان انجام سرچ بانکهای جهانی در مراکز پذیره نویسی و انباشت درخواستها در شبکه ملی و در نتیجه تاخیر در ارائه نتیجه سرچ به بیماران